Cycle
« Analyse de données de séquençage à haut-débit »

Module 1/5: Analyses ADN

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Module 1/5: Analyses ADN

- NGS Introduction
- Reads Quality Control
- Reads Cleaning
- Aligning reads on reference → *Hélène Touzet*
- Alignment parameters → *Hélène Touzet*
- Reads duplicates
- Assembly → *Hélène Touzet*

This training course was initially developed by Sophie Gallina (CNRS). Additional contributions were made by Chadi Saad (CHU Lille), Hélène Touzet (CNRS) and Pierre Pericard (Univ. Lille)

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- NGS Introduction
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Module 1/5: Analyses ADN

- NGS Introduction

➔ What is NGS?

➔ Sequencers

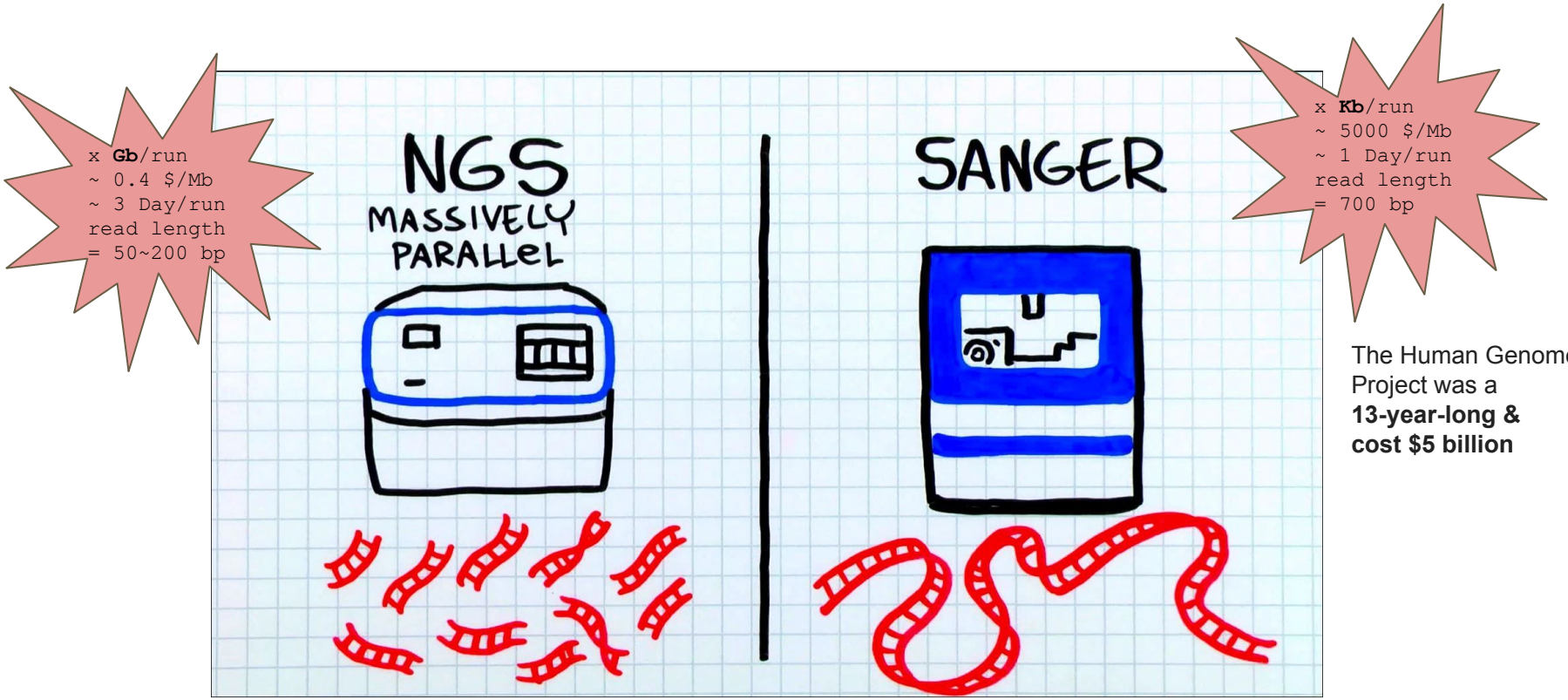
➔ Applications

➔ NGS workflow

➔ Output data

What is Next-Generation Sequencing (NGS)?

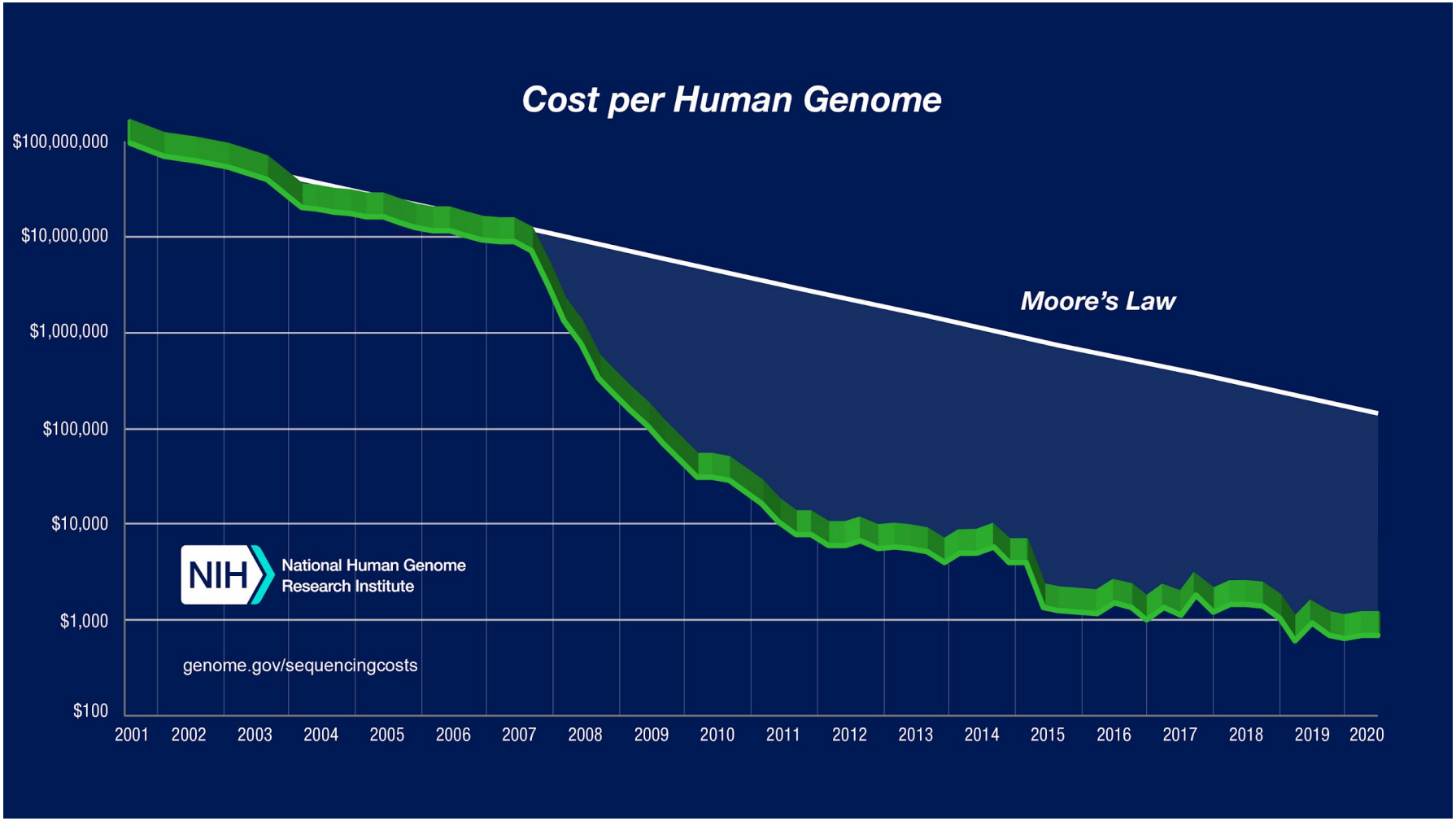
"Next-generation sequencing (NGS), also known as high-throughput sequencing, is the catch-all term used to describe a number of different modern sequencing technologies. These technologies allow for sequencing of DNA and RNA much more quickly and cheaply than the previously used Sanger sequencing, and as such revolutionised the study of genomics and molecular biology"



The Human Genome Project was a **13-year-long & cost $5 billion**

https://www.ebi.ac.uk/training/online/course/ebi-next-generation-sequencing-practical-course/what-you-will-learn/what-next-generation-dna-
https://www.thermofisher.com/blog/behindthebench/when-do-i-use-sanger-sequencing-vs-ngs-seq-it-out-7/

What is Next-Generation Sequencing (NGS)?



What is Next-Generation Sequencing (NGS)?

Illumina sequencing

Illumina sequencing works by simultaneously identifying DNA bases, as each base emits a unique fluorescent signal, and adding them to a nucleic acid chain

Ion Torrent: Proton / PGM sequencing (thermofisher)

Ion Torrent sequencing measures the direct release of H+ (protons) from the incorporation of individual bases by DNA polymerase and therefore differs from the previous two methods as it does not measure light.

https://www.ebi.ac.uk/training/online/course/ebi-next-generation-sequencing-practical-course/what-you-will-learn/what-next-generation-dna-

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Roche 454 pyrosequencing

https://www.ebi.ac.uk/training/online/course/ebi-next-generation-sequencing-practical-course/what-you-will-learn/what-next-generation-dna-

Sequencers – Illumina

illumina®

	Benchtop Sequencers			Production-Scale Sequencers	
	 iSeq 100	 MiniSeq	 MiSeq Series	 NextSeq 550 Series	 NextSeq 1000 & 2000
Popular Applications & Methods	Key Application	Key Application	Key Application	Key Application	Key Application
Large Whole-Genome Sequencing (human, plant, animal)					
Small Whole-Genome Sequencing (microbe, virus)	●	●	●	●	●
Exome & Large Panel Sequencing (enrichment-based)				●	●
Targeted Gene Sequencing (amplicon-based, gene panel)	●	●	●	●	●
Single-Cell Profiling (scRNA-Seq, scDNA-Seq, oligo tagging assays)				●	●
Transcriptome Sequencing (total RNA-Seq, mRNA-Seq, gene expression profiling)				●	●
Targeted Gene Expression Profiling	●	●	●	●	●
miRNA & Small RNA Analysis	●	●	●	●	●
DNA-Protein Interaction Analysis (ChIP-Seq)			●	●	●
Methylation Sequencing				●	●
16S Metagenomic Sequencing		●	●	●	●
Metagenomic Profiling (shotgun metagenomics, metatranscriptomics)				●	●
Cell-Free Sequencing & Liquid Biopsy Analysis				●	●
Run Time	9.5–19 hrs	4–24 hours	4–55 hours	12–30 hours	11-48 hours
Maximum Output	1.2 Gb	7.5 Gb	15 Gb	120 Gb	330 Gb*
Maximum Reads Per Run	4 million	25 million	25 million †	400 million	1.1 billion*
Maximum Read Length	2 × 150 bp	2 × 150 bp	2 × 300 bp	2 × 150 bp	2 × 150 bp

https://www.illumina.com/systems/sequencing-platforms.html

Sequencers – Illumina

illumina®

Benchtop Sequencers	Production-Scale Sequencers	
		
	NextSeq 550 Series	NextSeq 1000 & 2000
Popular Applications & Methods	Key Application	Key Application
Large Whole-Genome Sequencing (human, plant, animal)		
Small Whole-Genome Sequencing (microbe, virus)	●	●
Exome & Large Panel Sequencing (enrichment-based)	●	●
Targeted Gene Sequencing (amplicon-based, gene panel)	●	●
Single-Cell Profiling (scRNA-Seq, scDNA-Seq, oligo tagging assays)	●	●
Transcriptome Sequencing (total RNA-Seq, mRNA-Seq, gene expression profiling)	●	●
Chromatin Analysis (ATAC-Seq, ChIP-Seq)	●	●
Methylation Sequencing	●	●
Metagenomic Profiling (shotgun metagenomics, metatranscriptomics)	●	●
Cell-Free Sequencing & Liquid Biopsy Analysis	●	●
Run Time	12–30 hours	11-48 hours
Maximum Output	120 Gb	330 Gb*
Maximum Reads Per Run	400 million	1.1 billion*
Maximum Read Length	2 × 150 bp	2 × 150 bp

	NovaSeq 6000
	
	Key Application
Large Whole-Genome Sequencing (human, plant, animal)	●
Small Whole-Genome Sequencing (microbe, virus)	●
Exome & Large Panel Sequencing (enrichment-based)	●
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Single-Cell Profiling (scRNA-Seq, scDNA-Seq, oligo tagging assays)	●
Transcriptome Sequencing (total RNA-Seq, mRNA-Seq, gene expression profiling)	●
Chromatin Analysis (ATAC-Seq, ChIP-Seq)	●
Methylation Sequencing	●
Metagenomic Profiling (shotgun metagenomics, metatranscriptomics)	●
Cell-Free Sequencing & Liquid Biopsy Analysis	●
~13 - 38 hours (dual SP flow cells) ~13–25 hours (dual S1 flow cells) ~16–36 hours (dual S2 flow cells) ~44 hours (dual S4 flow cells)	
6000 Gb	
20 billion	
2 × 250**	

https://www.illumina.com/systems/sequencing-platforms.html

Sequencers – Illumina (pre-2020)

illumina®

NextSeq Series

HiSeq Series

HiSeq X Series‡

NovaSeq 6000 System

Popular Applications & Methods	Key Application	Key Application	Key Application	Key Application
Large Whole-Genome Sequencing (human, plant, animal)	●	●	●	●
Small Whole-Genome Sequencing (microbe, virus)	●	●		●
Exome Sequencing	●	●		●
Targeted Gene Sequencing (amplicon, gene panel)	●	●		●
Whole-Transcriptome Sequencing	●	●		●
Gene Expression Profiling with mRNA-Seq	●	●		●
miRNA & Small RNA Analysis	●	●		●
DNA-Protein Interaction Analysis	●	●		●
Methylation Sequencing	●	●		●
Shotgun Metagenomics	●	●		●

Optimized NGS Sample Tracking and Workflows

See how BaseSpace Clarity LIMS (Laboratory Information Management System) enabled this large genomics lab to standardize lab procedures and cope with increasing sample volumes from diverse clients.

Read Case Study ›

Run Time	12–30 hours	< 1–3.5 days (HiSeq 3000/HiSeq 4000) 7 hours–6 days (HiSeq 2500)	< 3 days	16–36 hours (Dual S2 flow cells) 44 hours (Dual S2 flow cells)
Maximum Output	120 Gb	1500 Gb	1800 Gb	6000 Gb§
Maximum Reads Per Run	400 million	5 billion	6 billion	20 billion**
Maximum Read Length	2 × 150 bp	2 × 150 bp	2 × 150 bp	2 × 150 bp

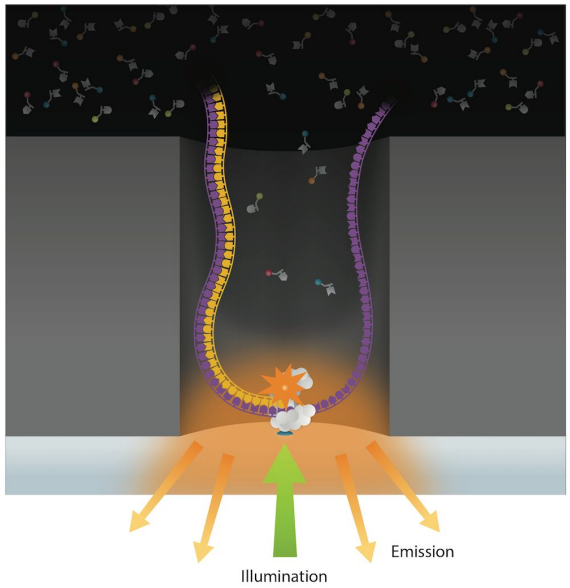
https://www.illumina.com/systems/sequencing-platforms.html

Sequencers – Thermo Fisher Scientific

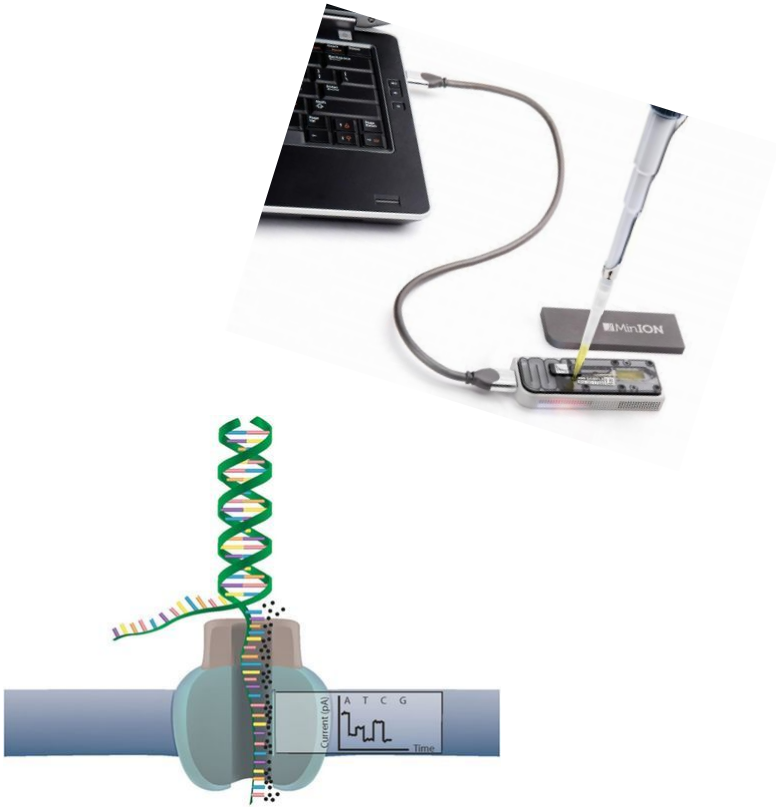
Plateformes de séquençage	 Système Ion PGM™ pour le séquençage de nouvelle génération	 Système Ion S5™ pour le séquençage de nouvelle génération	 Système Ion S5™ XL pour le séquençage de nouvelle génération
Avantages	**Évolutivité** : de 30 Mo à 2 Go **Rapidité** : séquençage exécuté en 2 à 7 heures, selon la longueur de lecture et la sortie par la puce	**Simplicité** : solutions de flux de travaux automatisé, de la préparation des échantillons à l'analyse **Évolutivité** : de 600 Mo à 15 Go **Rapidité** : séquençage effectué en 2,5 à 4 heures (quelle que soit la sortie par la puce)	**Simplicité** : solutions de flux de travaux automatisé, de la préparation des échantillons à l'analyse **Évolutivité** : de 600 Mo à 15 Go **Rapidité** : de l'ADN aux données en 24 heures
Applications de séquençage	ARN ciblé ADN ciblé Microbien	ARN ciblé ADN ciblé Microbien Transcriptome Exome Séquençage de l'ARN	ARN ciblé ADN ciblé Microbien Transcriptome Exome Séquençage de l'ARN

https://www.thermofisher.com/fr/fr/home/life-science/sequencing/sequencing-technology-solutions.html

Third-generation sequencing



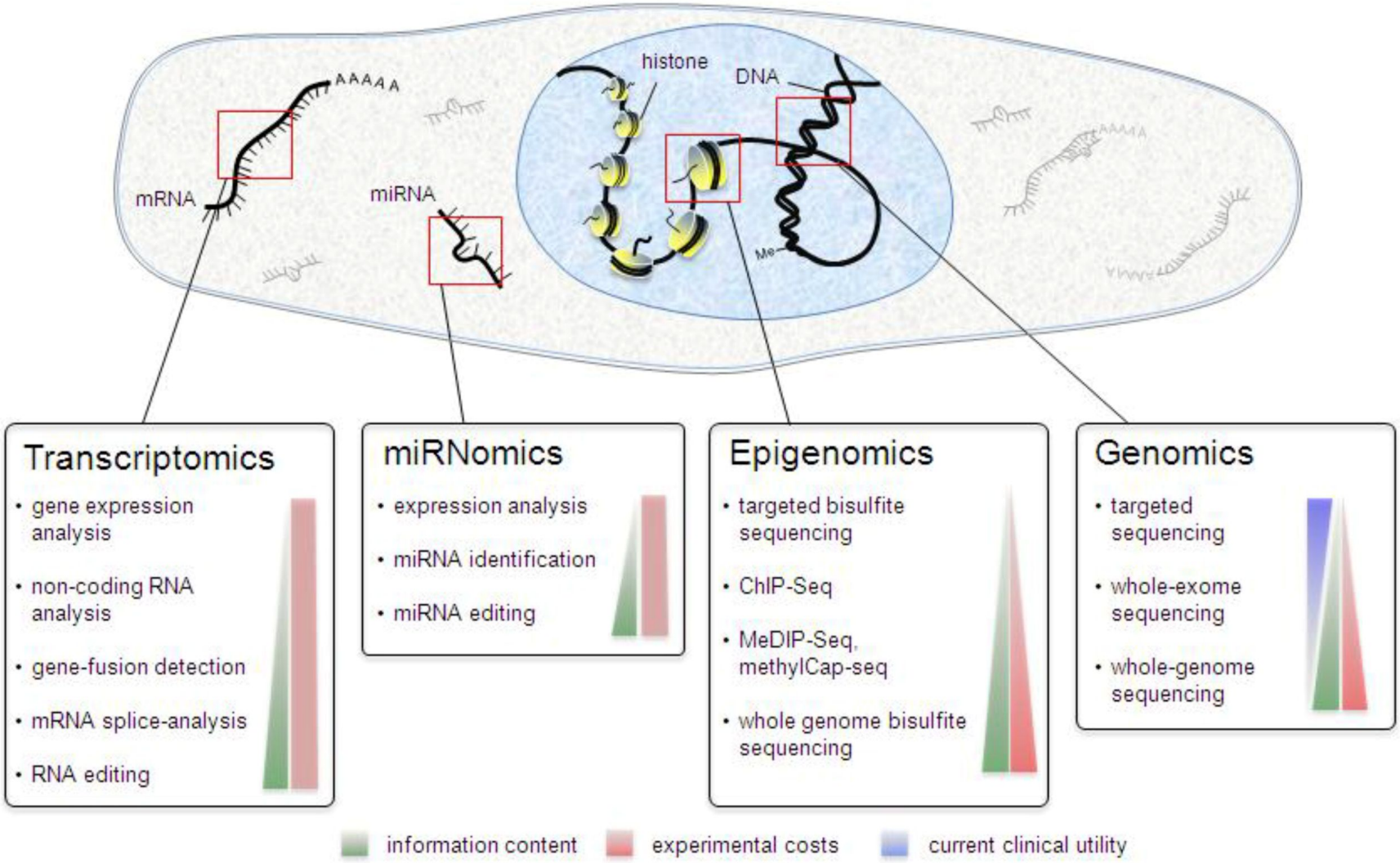
PacBio Sequencing



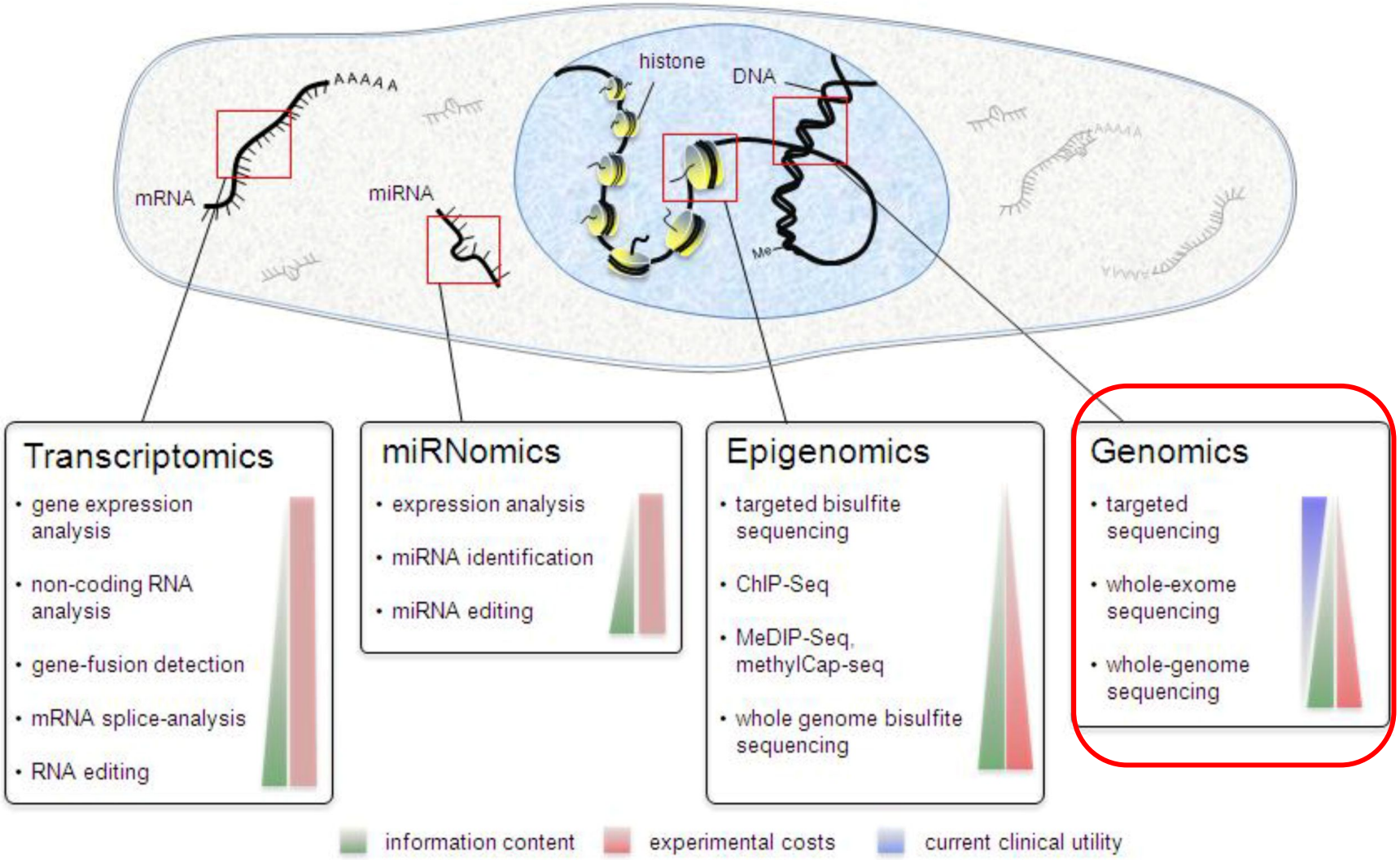
Nanopore technology

- much longer reads (> Kb)
- error rate (~ 10 → 40 %)

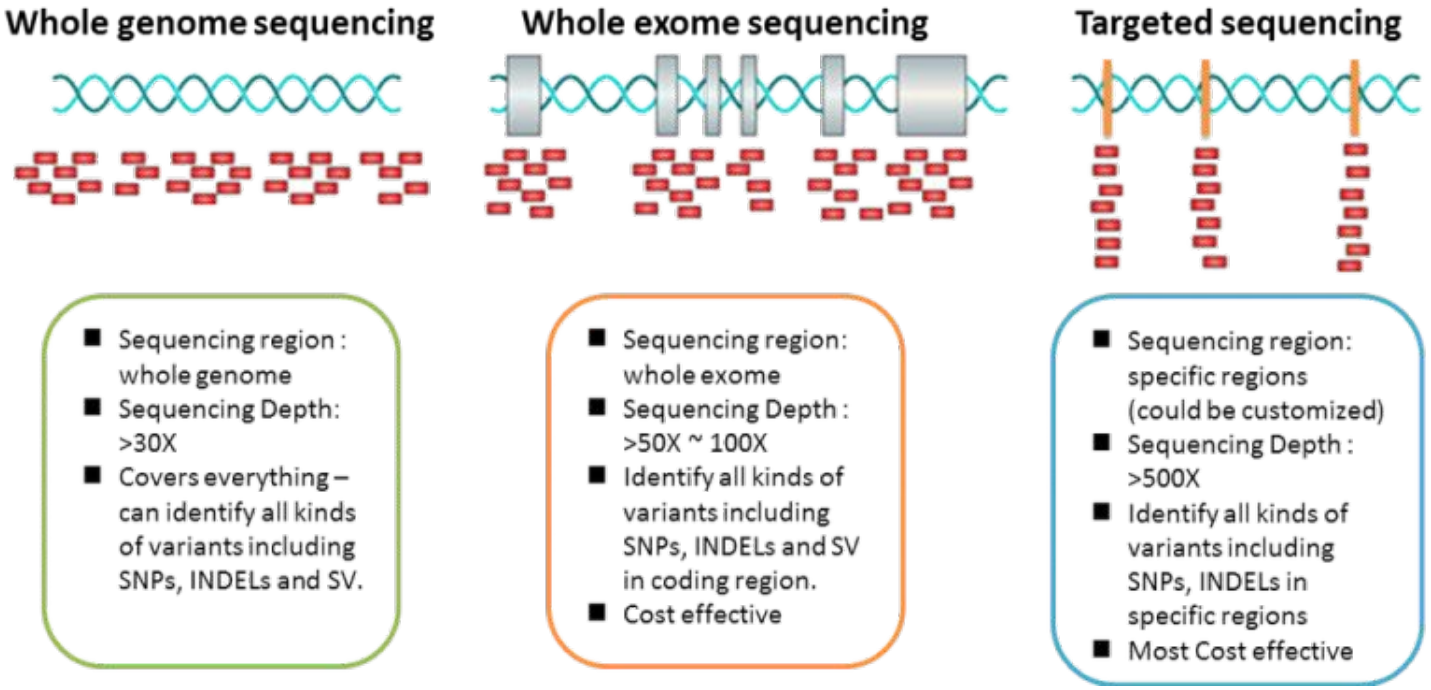
Applications



Applications



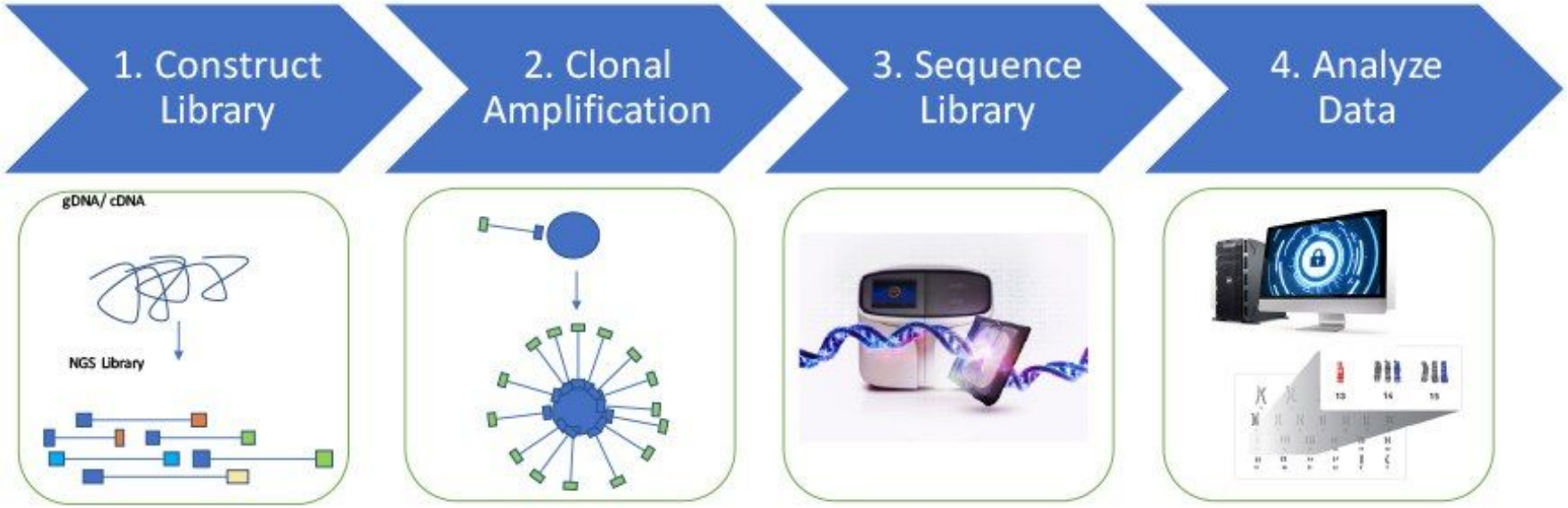
Applications : genomics (DNA-seq)



- Targeted sequencing : rapid and cost-effective way to detect known and novel variants in selected sets of genes or genomic regions
- Whole exome sequencing : sequencing all of the protein-coding regions of genes in a genome (applications : discover rare-variants, adjacent splice-sites,...)
- Whole genome sequencing : alterations in regulatory sequences and non-coding regions, chromosomal rearrangements, ...

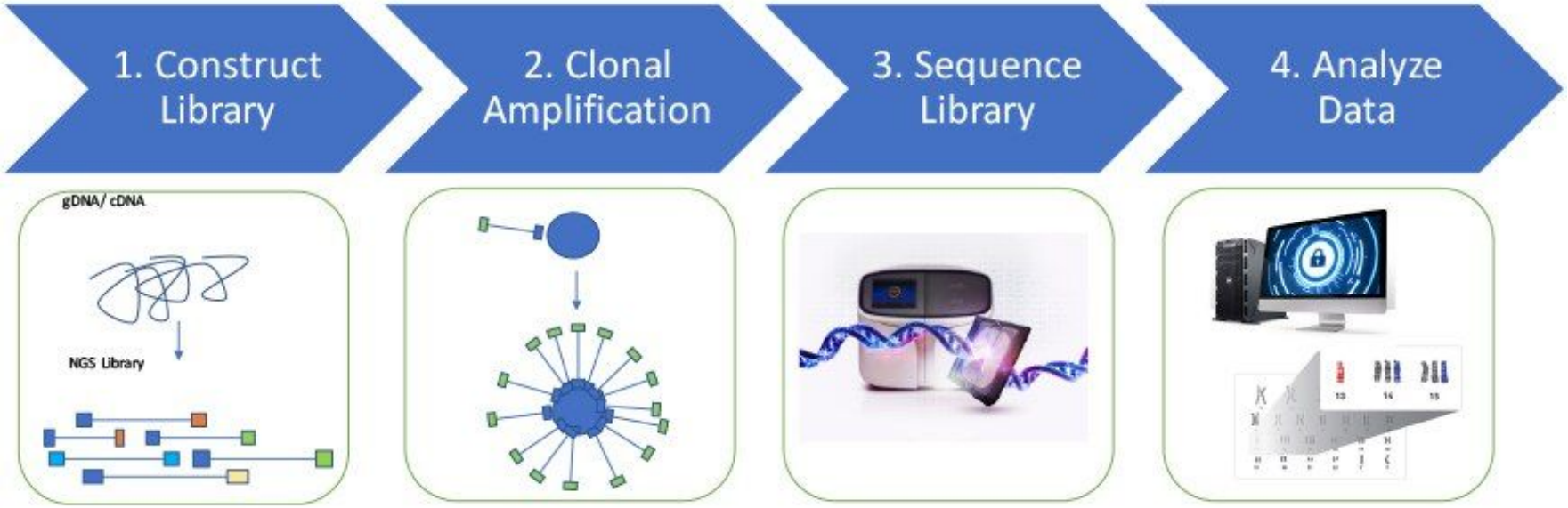
http://www.genomesop.com/somatic-mutations/

NGS workflow



https://www.thermofisher.com/fr/fr/home/life-science/sequencing/sequencing-education/next-generation-sequencing-basics/what-is-next-generation-sequencing.html

NGS workflow



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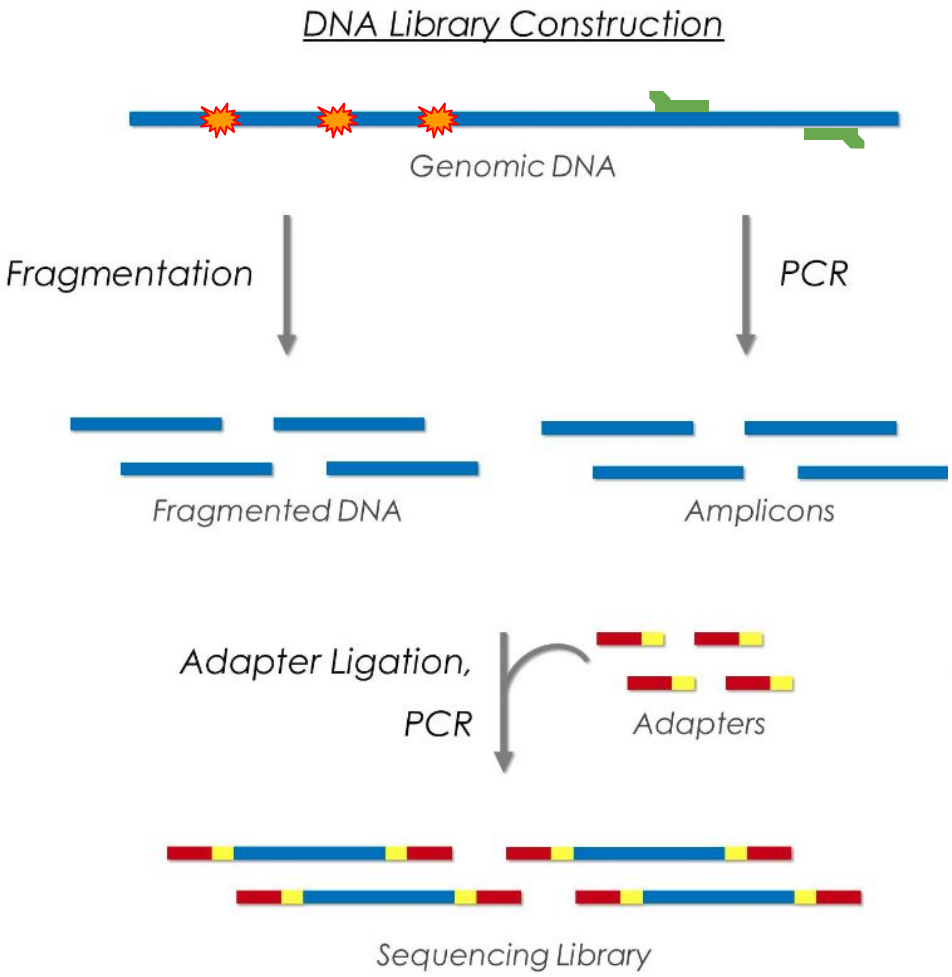
Library construction

A sequencing “library” must be created from the sample. The DNA (or cDNA) sample is processed into relatively short double-stranded fragments (100–800 bp)



source : https://www.biocompare.com/Molecular-Biology/9187-Next-Generation-Sequencing/

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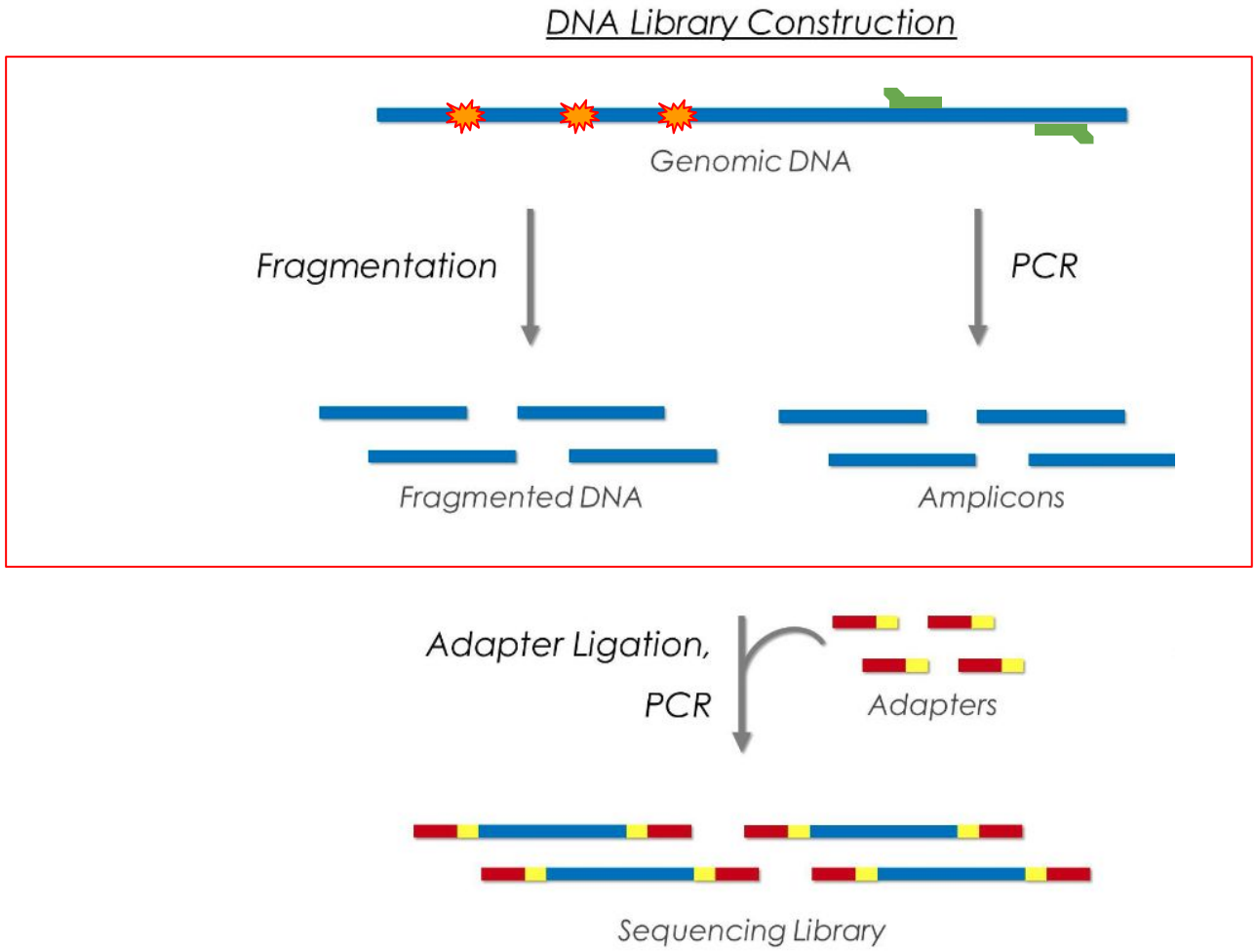


source : https://www.biocompare.com/Molecular-Biology/9187-Next-Generation-Sequencing/

Library construction

Targeted sequencing : enrichment methods

Effective in enrichment and specificity

Simple and fast protocol

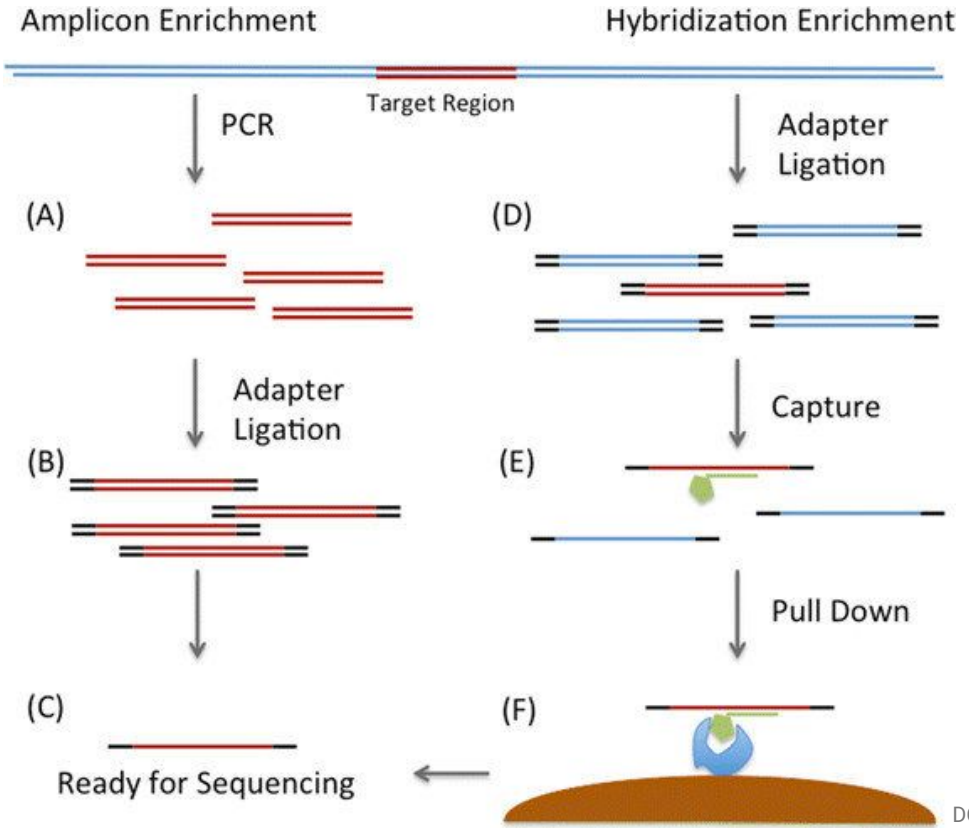
Target from Kb to Mb

Low DNA input (100 ng)

HaloPlex

AmpliSeq

...



Effective in enrichment and specificity

Complex procedure

Larger gene panels

Higher DNA input (>1 µg)

Agilent's SureSelect

Roche/Nimbelgen's SeqCap

Illumina's TruSeq and Nextera

...

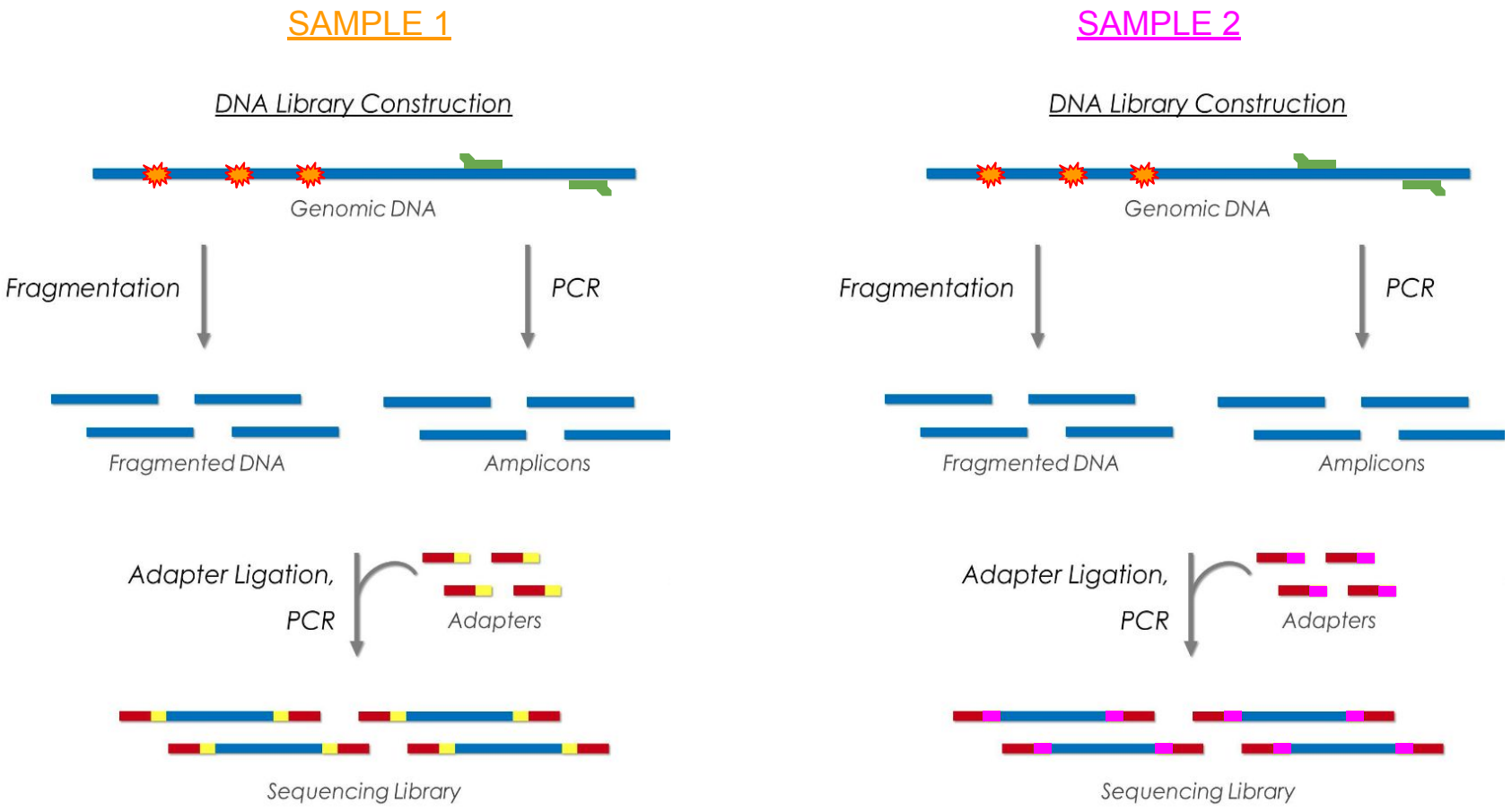
DOI: 10.1186/s13075-014-0490-4

The **BED** format is a text file format used to store genomic regions as coordinates and associated annotations

```
chr7    127471196    127472363
chr7    127472363    127473530
chr7    127473530    127474697
```

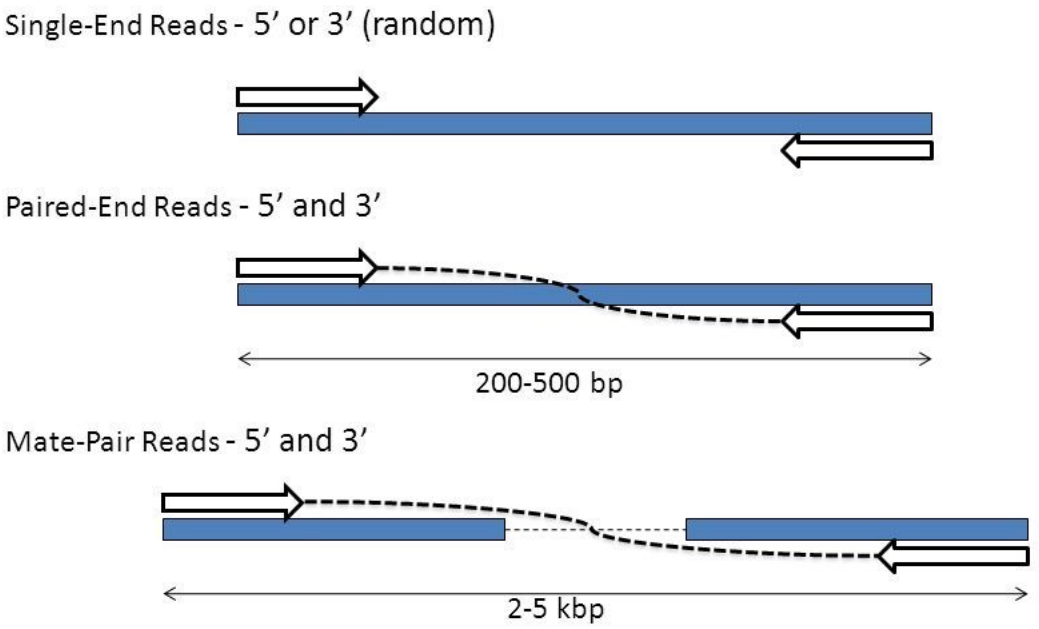
Library construction

Multiplex sequencing using DNA barcoding



Single-end vs paired-end

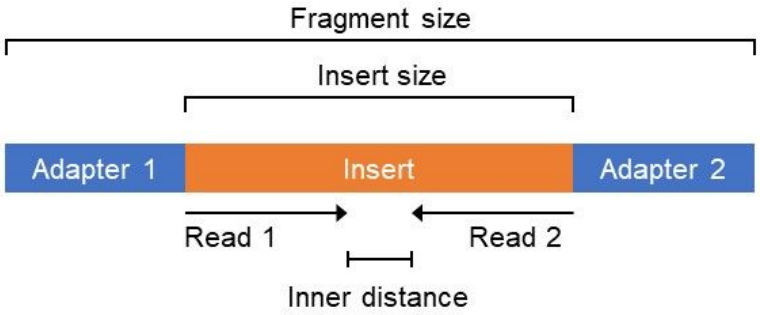
- **Single-End Read:** When sequencing process only occurs in 1 direction
- **Paired-End Read:** When sequencing process occurs in both directions
- **Mate-pair Read:** Short fragments consisting of two segments that originally had a separation of several kilobases in the genome.

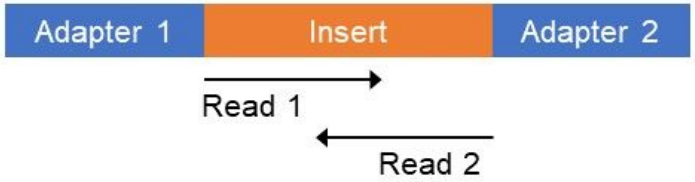


source: http://slideplayer.com/slide/7847747/25/images/7/Types+of+Sequencing+Libraries.jpg

Paired-end

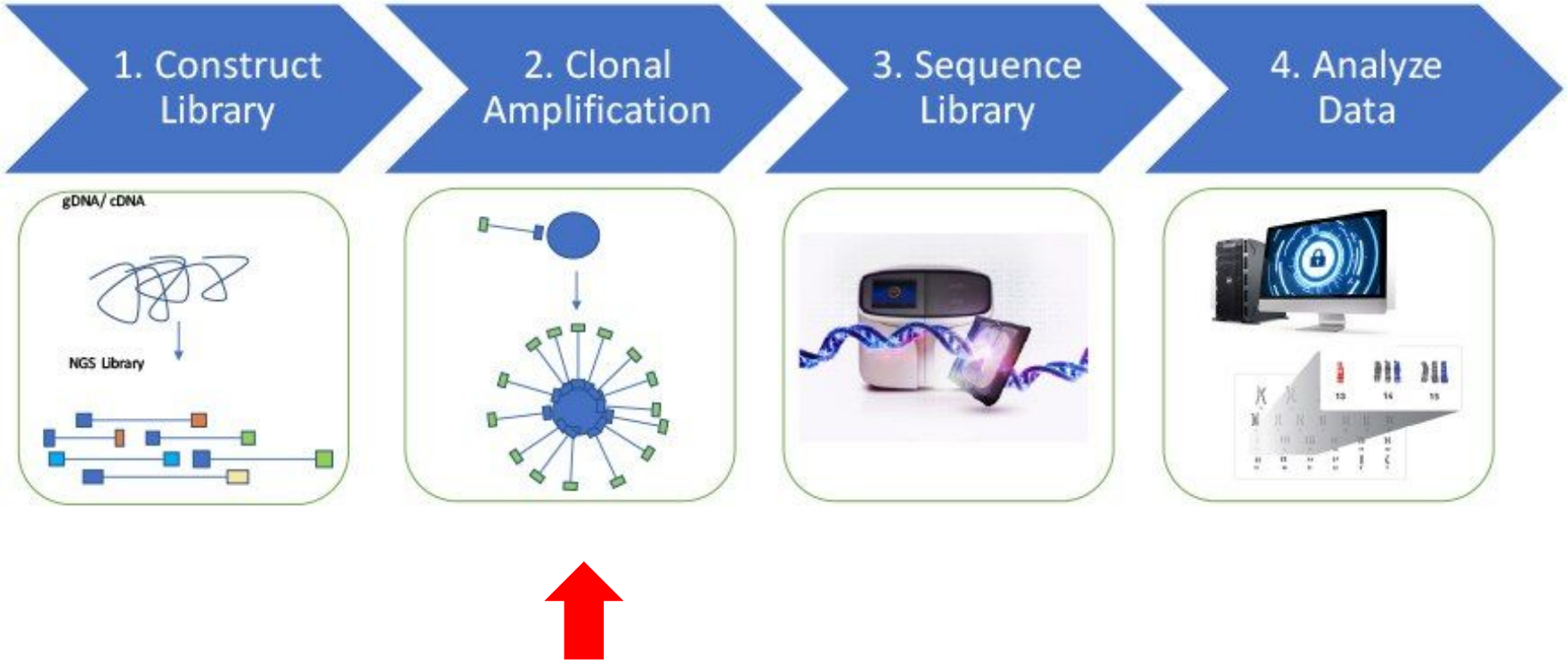
➢ **The insert size** is the size of the piece of DNA of interest, without the adapters.





https://www.badgrammargoodsyntax.com/compbio/2018/3/5/compbio-020-reads-fragments-and-inserts-what-you-need-to-know-for-understanding-your-sequencing-data-1

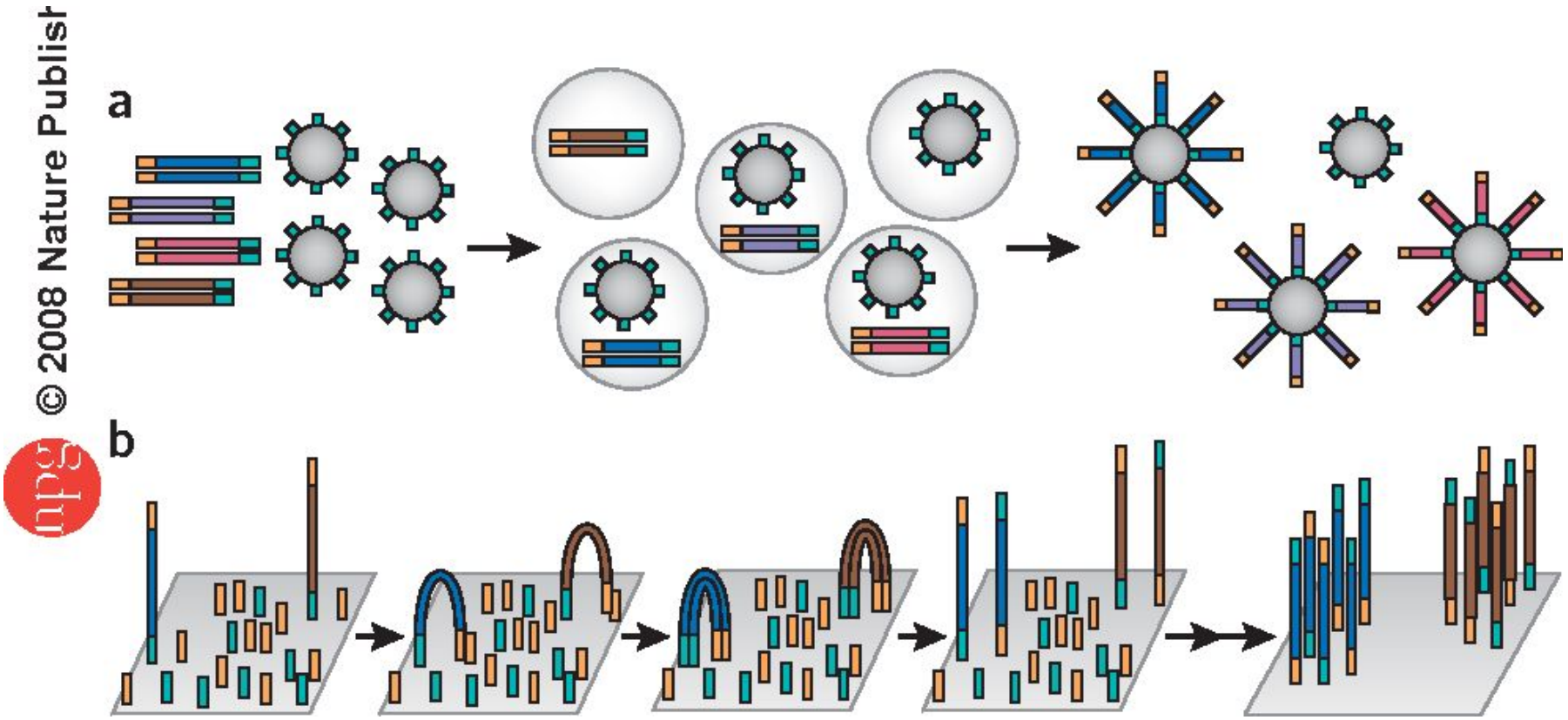
NGS workflow



https://www.thermofisher.com/fr/fr/home/life-science/sequencing/sequencing-education/next-generation-sequencing-basics/what-is-next-generation-sequencing.html

Clonal amplification

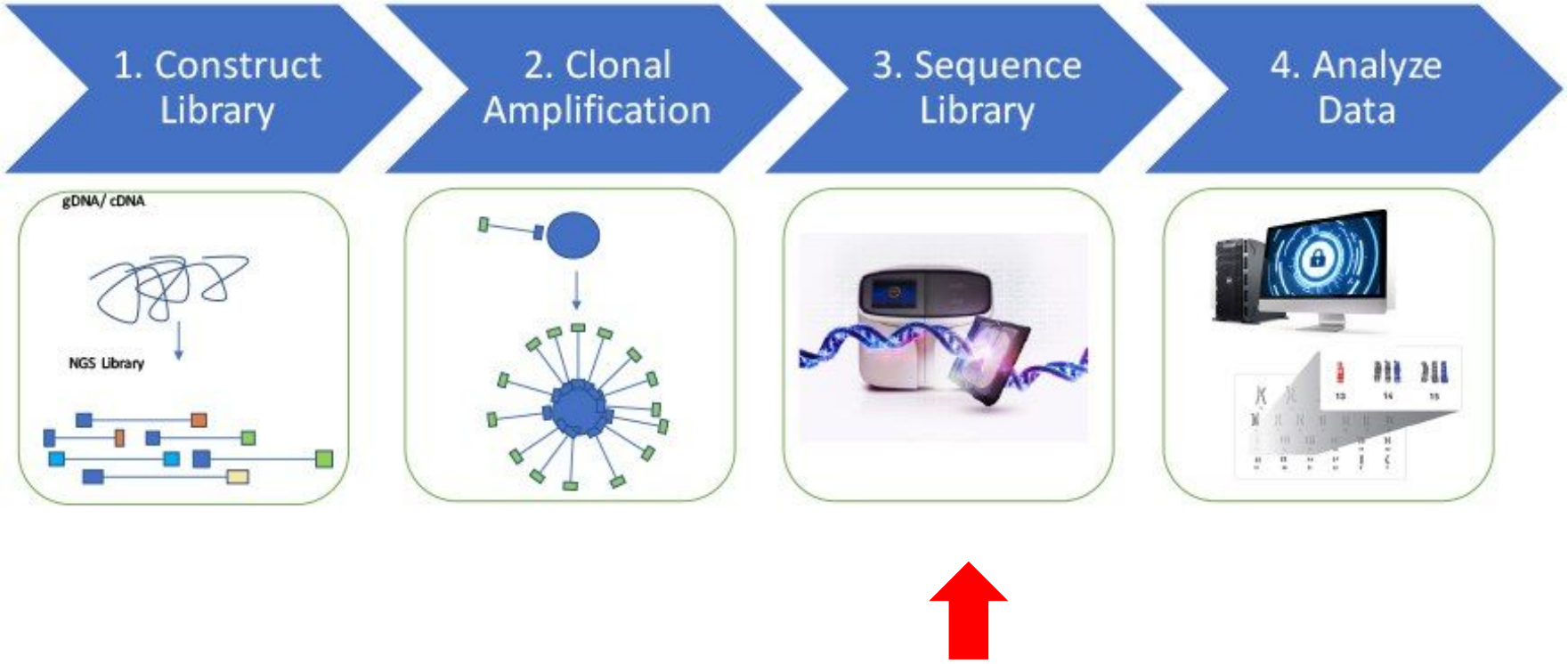
Prior to sequencing, the DNA library must be attached to a solid surface and clonally amplified to increase the signal that can be detected from each target during sequencing.



(a) thermofisher platforms rely on emulsion to amplify clonal sequencing features. (b) The Illumina technology relies on bridge PCR21,22 (aka 'cluster PCR') to amplify clonal sequencing features.

Nature Biotechnology 2008, DOI:10.1038/nbt1486

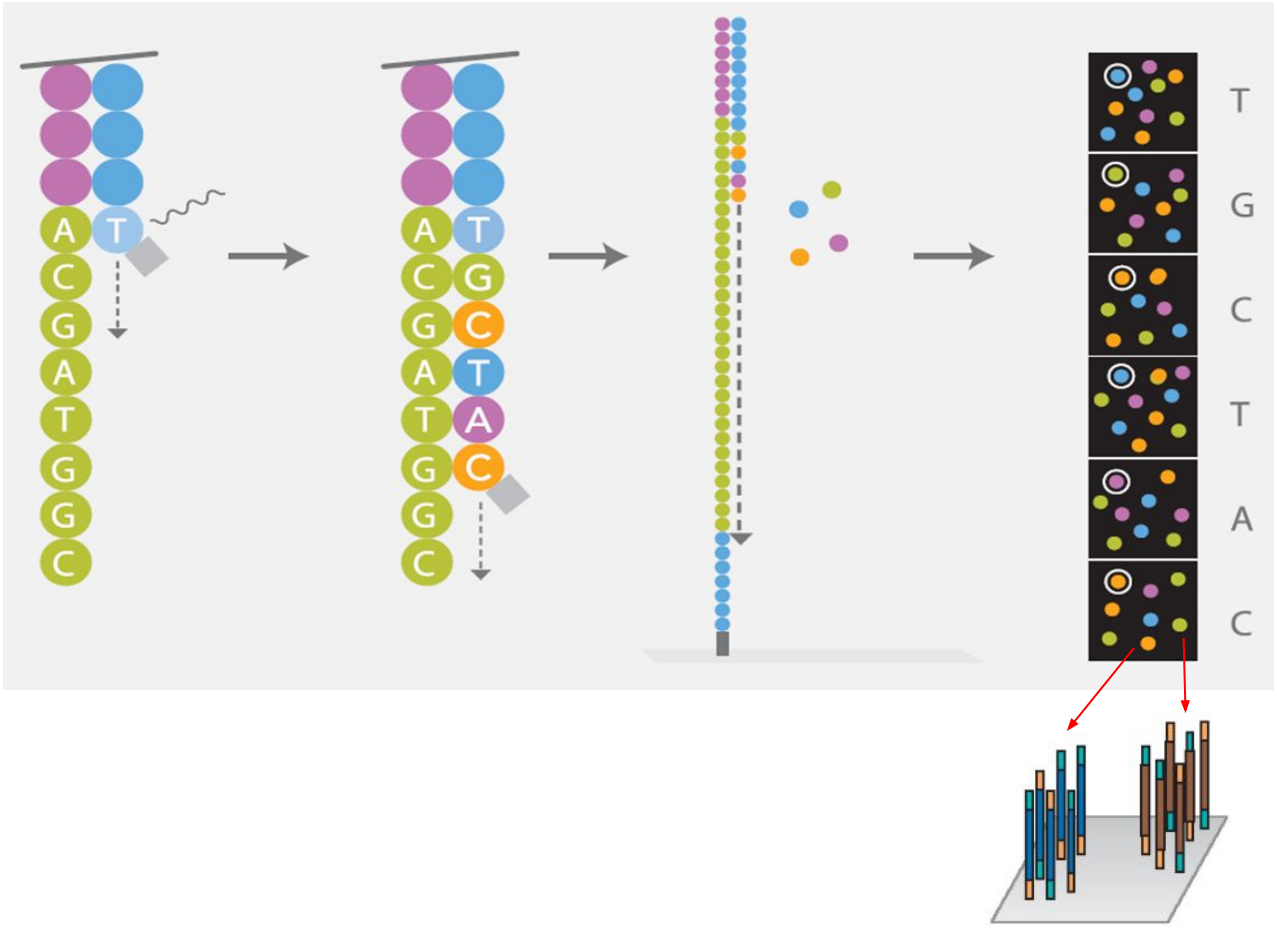
NGS workflow

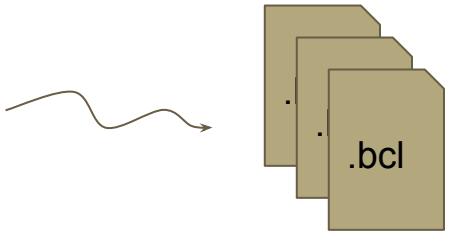


https://www.thermofisher.com/fr/fr/home/life-science/sequencing/sequencing-education/next-generation-sequencing-basics/what-is-next-generation-sequencing.html

Sequencing

Illumina technology

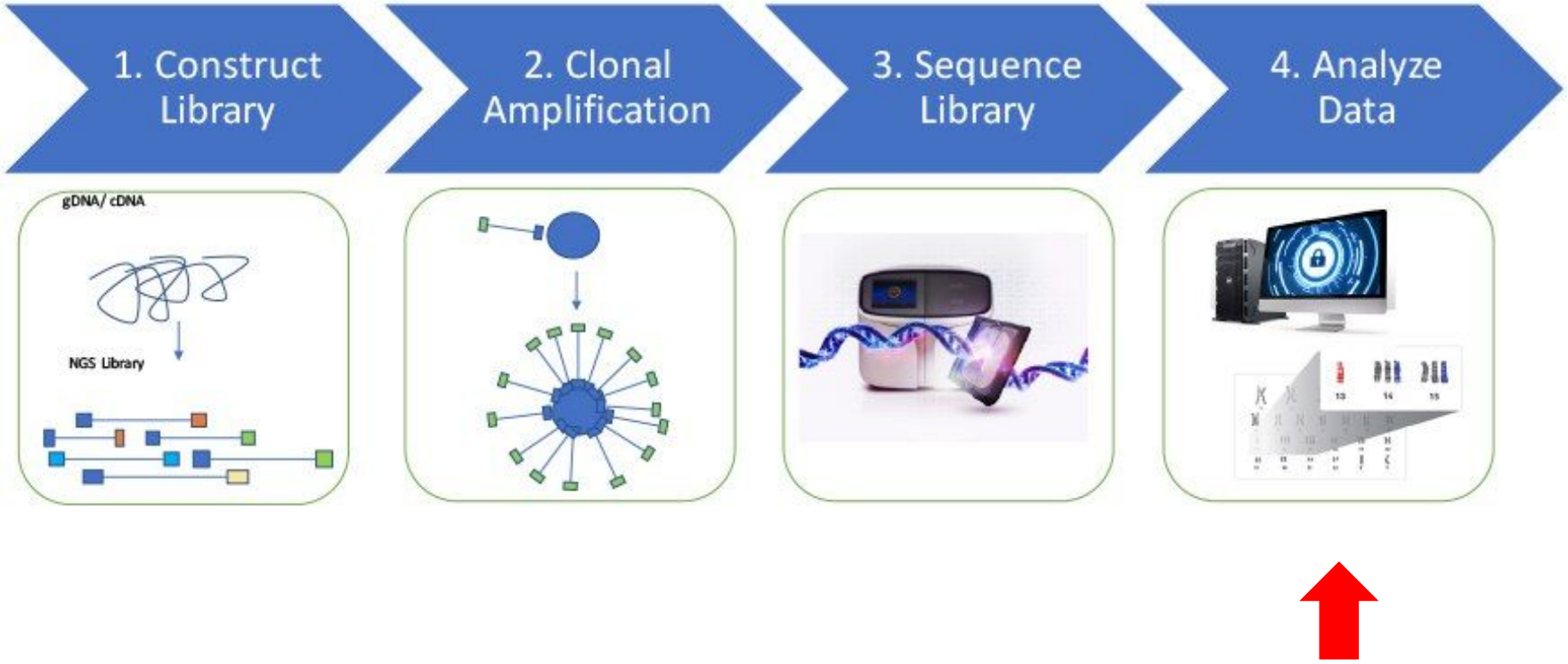




Raw data

https://www.cegat.de/en/services/next-generation-sequencing/

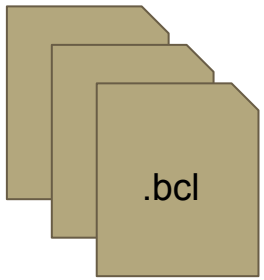
NGS workflow



https://www.thermofisher.com/fr/fr/home/life-science/sequencing/sequencing-education/next-generation-sequencing-basics/what-is-next-generation-sequencing.html

Data analyses

Extracting reads, Demultiplexing



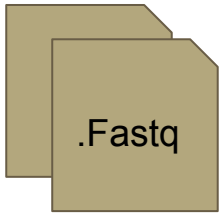
+

Sample Sheet

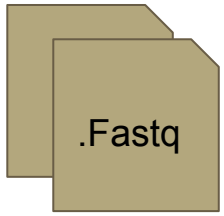
[Header]							
IEMFileVersion	4						
Experiment Name	Project1						
Date	4/16/2016						
Workflow	GenerateFASTQ						
Application	NextSeq FASTQ Only						
Assay	TruSeq LT						
Description							
Chemistry	Default						
[Reads]							
151							
151							
[Settings]							
Adapter	AGATCGGAAGAGCACACGTCTGAACTCCAGTCA						
AdapterRead2	AGATCGGAAGAGCGTCGTGTAGGGAAAGAGTGT						
[Data]							
Sample_ID	Sample_Name	Sample_Plate	Sample_Well	I7_Index_ID	index	Sample_Project	Description
Sample_1				A002	CGATGT		
Sample_2				A004	TGACCA		
Sample_3				A005	ACAGTG		
Sample_4				A006	GCCAAT		

bcl2fastq

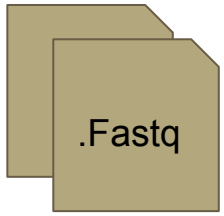
sample 1



sample 2

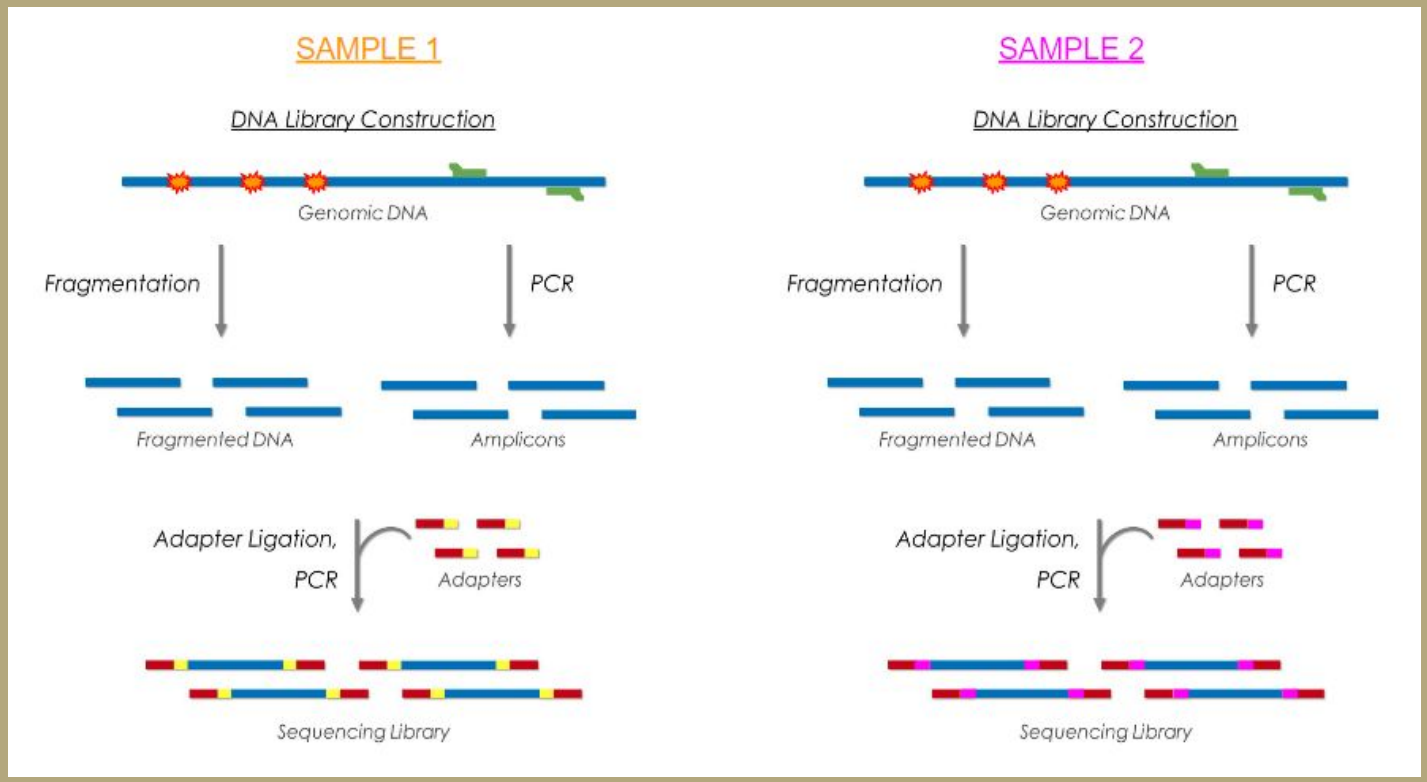


sample 3



Data analyses

Extracting reads, Demultiplexing



sample 1

.Fastq

sample 2

.Fastq

sample 3

.Fastq

Fastq files (Paired-end)

2 files : R1, R2

Reads1.fq

```
@ERR229776.100000840
CTAGGAAGCGTAGTCCTGGGGTCATCTCTCCTATTAATACTGTTGGGGAATGTTTAGTA
+
BAEEAGEED96EHFE@BF><>EAAC;EBH<K<6:HJGFFHBC>DDIKG4AIHFFD@0/=
@ERR229776.100020365
CATTATTTCATAGTAGCCAAAAAGTGGAAACAGTCAAAATATCCGTCAGTGAATTGACC
+
1.*/./&((&3=;B@F860C>@51(3:).6GG-68C*:CG)#B4/=HDJ6;79)<@C/
@ERR229776.100104918
TATTTCTGGAATTTTCCATTTAATATTTTCAGACTGCAGTTGACTGCGGGTAACTGAAA
+
CEEEEEFEDAEGGGFDHGFFHGIHHHIIIIGKHBKJJIGHFHKILJKLEJLJJIFJMJK
```

Reads2.fq

```
@ERR229776.100000840
TTCTGGTCAGTAAGACCTCAAAAGGTTAAATACTAGCGATTTACACACCTTAAATGATT
+
CFIEEG@FFFGKFJHJ>HHKLLJIIJILLJIILJHKAKJKKJJJJJJLMKJMKJJJJKJ
@ERR229776.100020365
CCTAAAATGGTGTGTTTTCGTATATTCACAATGCTGTGGAACCATCACCACTATCTGAT
+
4B@EDFF=(/CHBHEHCE6@ED8E@@I6HJB6E:6%@C46FFIBGCIGKD,DN=CBBE@
@ERR229776.100104918
TCTTTCTTTTGTTTTTTTTTCTGAGATGTCTTTTGTTTTTGTTCTGAGGTCTTGTTATG
+
CFIGGGKHHHFHHFIJIIIJKLIIHJIIIKLJKKIJKLLKJFJJMHJJLFJMJIKKJJJ
```

1 interleaved paired file

Reads.fq

```
@SRR531199.1 ILLUMINA_0130:3:1101:1249:1993 length=101
TTTTCAGAGTAGTTGGTACCCAATTGGAAGATGTGACCCACTTCGATACCGCGCTTGAG
+
dfffffffdffeffdadfffeeefdeffeffeffffffffddeefeYdfefefe[e
@SRR531199.1 ILLUMINA_0130:3:1101:1249:1993 length=99
ANNNNNNCTTCGGTATNAACTGGGGNNNNGATGTTGAACTGGGTAAAGTCGAAGATCTG
+
BBBBBBSZTUVWO]YB_[cbabbWBBBBSVVUUgggadcdedbedcddfffdegeggef
@SRR531199.2 ILLUMINA_0130:3:1101:1463:1964 length=101
NTGAGTAGCTCAATGCGCTGACGCCAATAGCTATACCAACGACTGGCCAGATTATGTTT
+
BXSSRU[X[Wcc_ccccccccccc_ccccccccc_ccccccccccccccccccccc
@SRR531199.2 ILLUMINA_0130:3:1101:1463:1964 length=99
AAGTGACCCATCGCGATAAAGTGCTGCGCAGTAAANAGCANCTGTTNGATGCTGGCTTA
+
ggggggggggggggggggfgfgggggggggggg^BbbbaBbbaZ]BZ[ccccfggggg
@SRR531199.3 ILLUMINA_0130:3:1101:1366:1970 length=101
NAAGTCGCGGCGACCCCTATCGTGGCTTTCGGCGTACGCCATTTCAATGCGGCCGCCGC
+
B[[X[YY[YVcc_cccc_cc__________[[[V[^^^^^V[[]SXWUX[\\]]Z^^^B
@SRR531199.3 ILLUMINA_0130:3:1101:1366:1970 length=99
TGGTCAATACAAGCCGCAATACCTGCATCATGCGGNGGAANAATTTGCGCGCCGTTTTC
+
ggfegggggggdeggggfgcgggaggggggggega^Bb`^]B[Y[[[Zffffh_afeefe
```

Fastq file format

1. Identifier
2. Sequence
4. Quality scores *(as ASCII chars)*

READ

```
@SRR062641.6751359
CGCCCGGCCAATCATTGTGGTTTTAAGTCACTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCT
+
CBLNPGJQQQJPPQPPQPQRGPPPPRRQQRPSPGRQQQRLRRRMEPQQPMJHQQEHKMMFIIRH?SIIHKNJIKRLJJIKHEABHIFGCGGEFCGDGDCE

@SRR062634.16249693
CTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCTCTGTGCACCCAGCATTGCCCAGAACAGGGC
+
ALKMOOOOPPQJQOPPPPPQPPPPPPPRJQRQQQQQRPQPRQQPFQSQQPRLIMHKSNRJQORMFELRPQNQRQJQRRPQQLIRKDMKQJPN8CFDGCCCB

@SRR062634.20060465
CTCCCAGCTTCCAACAGACCCTGTCCCAGCTCCCTCCAAGCTGAGTGTTGGCCTGATACCTACCAGTGGAGCGAGGGGAACCCGAGGACTGCCAAGGGCA
+
D?KMPQEPGCPQQNPQIQIGR@DPERQHEKBED=HCHG8EHFDCD6<329@<:69A<6,;<967>;=C:>AA8BBED######################
```

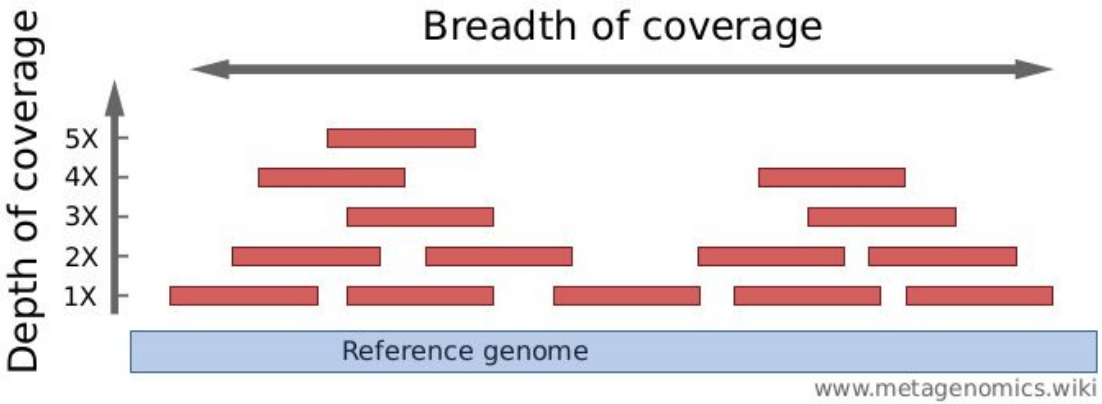
ASCII table:

Decimal	Hex	Char	Decimal	Hex	Char	Decimal	Hex	Char	Decimal	Hex	Char
0	0	[NULL]	32	20	[SPACE]	64	40	@	96	60	`
1	1	[START OF HEADING]	33	21	!	65	41	A	97	61	a
2	2	[START OF TEXT]	34	22	"	66	42	B	98	62	b
3	3	[END OF TEXT]	35	23	#	67	43	C	99	63	c
4	4	[END OF TRANSMISSION]	36	24	$	68	44	D	100	64	d
5	5	[ENQUIRY]	37	25	%	69	45	E	101	65	e
6	6	[ACKNOWLEDGE]	38	26	&	70	46	F	102	66	f
7	7	[BELL]	39	27	'	71	47	G	103	67	g
8	8	[BACKSPACE]	40	28	(	72	48	H	104	68	h
9	9	[HORIZONTAL TAB]	41	29	)	73	49	I	105	69	i
10	A	[LINE FEED]	42	2A	*	74	4A	J	106	6A	j
11	B	[VERTICAL TAB]	43	2B	+	75	4B	K	107	6B	k
12	C	[FORM FEED]	44	2C	,	76	4C	L	108	6C	l
13	D	[CARRIAGE RETURN]	45	2D	-	77	4D	M	109	6D	m
14	E	[SHIFT OUT]	46	2E	.	78	4E	N	110	6E	n
15	F	[SHIFT IN]	47	2F	/	79	4F	O	111	6F	o
16	10	[DATA LINK ESCAPE]	48	30	0	80	50	P	112	70	p
17	11	[DEVICE CONTROL 1]	49	31	1	81	51	Q	113	71	q
18	12	[DEVICE CONTROL 2]	50	32	2	82	52	R	114	72	r

Coverage and depth of coverage

- **Depth of coverage =** average number of reads covering a base (X)
  - Example: 30X for normal sample, 100X for tumor sample

- **(Breadth of) Coverage =** percentage of the targeted regions covered by at least X read
  - For example: 90% of a genome is covered at 1X depth; and still 40% is covered at 4X depth.



Source :
- Élodie Girard , 5ème Ecole de bioinformatique AVIESAN-IFB 2016 ,http://www.france-bioinformatique.fr/sites/default/files/V01_ITMO_2016_EG_from_fastq_to_mapping_1.pdf
- http://www.metagenomics.wiki/pdf/definition/coverage-read-depth

Module 1/5: Analyses ADN

- NGS Introduction
- Reads Quality Control
- Reads Cleaning
- Aligning reads on reference → *Hélène Touzet*
- Alignment parameters → *Hélène Touzet*
- Reads duplicates
- Assembly → *Hélène Touzet*

Reads quality

- Errors when reading bases
- Depends on sequencing technologie
- Error rate increases with read size

⇒ For each position in the read

- One base (A/T/C/G)
- One error probability

Phred Quality Score (for a base)

Phred quality scores ***Q*:** logarithmically related to the base-calling error probabilities $\mathcal{P}$

$$Q = -10 \log_{10} P$$

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10,000	99.99%
50	1 in 100,000	99.999%
60	1 in 1,000,000	99.9999%

source: https://en.wikipedia.org/wiki/Phred_quality_score

Quality score encoding

For history reasons, more than one coding convention

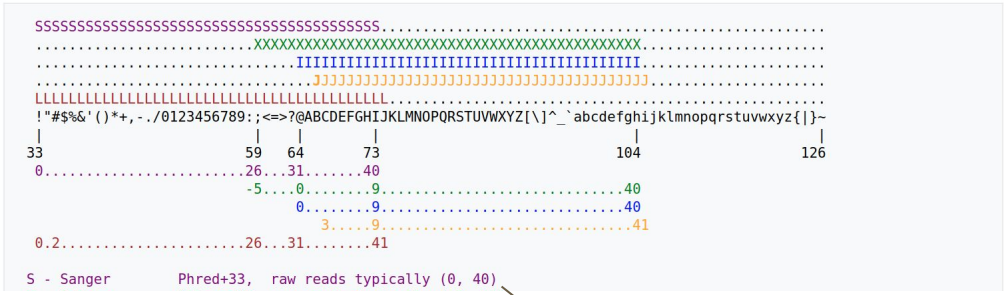
```
 SSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSSS.....................................................
 ..........................XXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXXX.....................
 ..............................IIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIIII...........................
 ................................JJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJJ.......................
 LLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLLL..................................................
 PPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPPP
 !"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHIJKLMNOPQRSTUVWXYZ[\]^_`abcdefghijklmnopqrstuvwxyz{|}~
 |                          |    |        |                              |                     |
33                          59   64       73                             104                   126
 0.........................26...31.......40
                            -5....0........9.............................40
                                 0........9.............................40
                                    3.....9.............................41
 0.2.......................26...31........41
 0...................20........30........40........50.....................................93
```

```
S - Sanger        Phred+33,  raw reads typically (0, 40)
X - Solexa        Solexa+64, raw reads typically (-5, 40)
I - Illumina 1.3+ Phred+64,  raw reads typically (0, 40)
J - Illumina 1.5+ Phred+64,  raw reads typically (3, 41)
    with 0=unused, 1=unused, 2=Read Segment Quality Control Indicator (bold)
    (Note: See discussion above).
L - Illumina 1.8+ Phred+33,  raw reads typically (0, 41)
P - PacBio        Phred+33,  HiFi reads typically (0, 93)
```

Source : https://fr.wikipedia.org/wiki/FASTQ

Galaxy : Always uses Sanger coding
=> conversion tool (*groomer*)

Example for score interpretation using sanger encoding



Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10,000	99.99%
50	1 in 100,000	99.999%
60	1 in 1,000,000	99.9999%

S - Sanger Phred+33

Bad : 0-19
Correct : 20-29
Good : 30-40

```
@SEQ:ID
ACTGTACGATCGATCGCATGCATCAGTACGTCGTACCAGAT
+
!"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHI
|                    |         |         |
0.........1.........2.........3.........4
01234567890123456789012345678901234567890
```

Goal: read cleaning

RAW

```
@SRR062641.6751359
CGCCCGGCCAATCATTGTGGTTTTAAGTCACTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCT
+
CBLNPGJQQQJPPQPPQPQRGPPPPRRQQRPSPGRQQQRLRRRMEPQQPMJHQQEHKMMFIIRH?SIIHKNJIKRLJJIKHEABHIFGCGGEFCGDGDCE
@SRR062634.16249693
CTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCTCTGTGCACCCAGCATTGCCCAGAACAGGGC
+
ALKMOOOOPPQJQOPPPPPQPPPPPPPRJQRQQQQQRPQPRQQPFQSQQPRLIMHKSNRJQORMFELRPQNQRQJQRRPQQLIRKDMKQJ#KNFDGCCCB
@SRR062634.20060465
CTCCCAGCTTCCAACAGACCCTGTCCCAGCTCCCTCCAAGCTGAGTGTTGGCCTGATACCTACCAGTGGAGCGAGGGGAACCCGAGGACTGCCAAGGGCA
+
D?KMPQEPGCPQQNPQIQIGR@DPERQHEKBE#HCHG8EHFDCD6<329@<:69A<6,;<967>;=C:>AA8BBED######################
@SRR062635.15516129
AAAAAAAAAAAAAAAAAAAAAAAAAAAAGGGGGCCCCCCTTTCCCCCCCGGGGGGGGGACAGGGGGGGTGTTCGGGCCCCGCGCCGCCCTTGACCACGG
+
EKLMPPPPPQQQQQQQQQQQQQQQQH#########################################################################
```

Clean

```
@SRR062641.6751359
CGCCCGGCCAATCATTGTGGTTTTAAGTCACTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCT
+
CBLNPGJQQQJPPQPPQPQRGPPPPRRQQRPSPGRQQQRLRRRMEPQQPMJHQQEHKMMFIIRH?SIIHKNJIKRLJJIKHEABHIFGCGGEFCGDGDCE
@SRR062634.16249693
CTAAGTTTGAGGCTATTTTGTTTTACAGCAAAAGCTAACTGATGCAGACAGGGACAAGTCAGTCTCATCTCTGTGCACCCAGCATTGCCCAGAACAGGGC
+
ALKMOOOOPPQJQOPPPPPQPPPPPPPRJQRQQQQQRPQPRQQPFQSQQPRLIMHKSNRJQORMFELRPQNQRQJQRRPQQLIRKDMKQJ#KNFDGCCCB
@SRR062634.20060465
CTCCCAGCTTCCAACAGACCCTGTCCCAGCTCCCTCCAAGCTGAG
+
D?KMPQEPGCPQQNPQIQIGR@DPERQHEKBE#HCHG8EHFDCD
```

Module 1/6: Analyses ADN

- NGS Introduction
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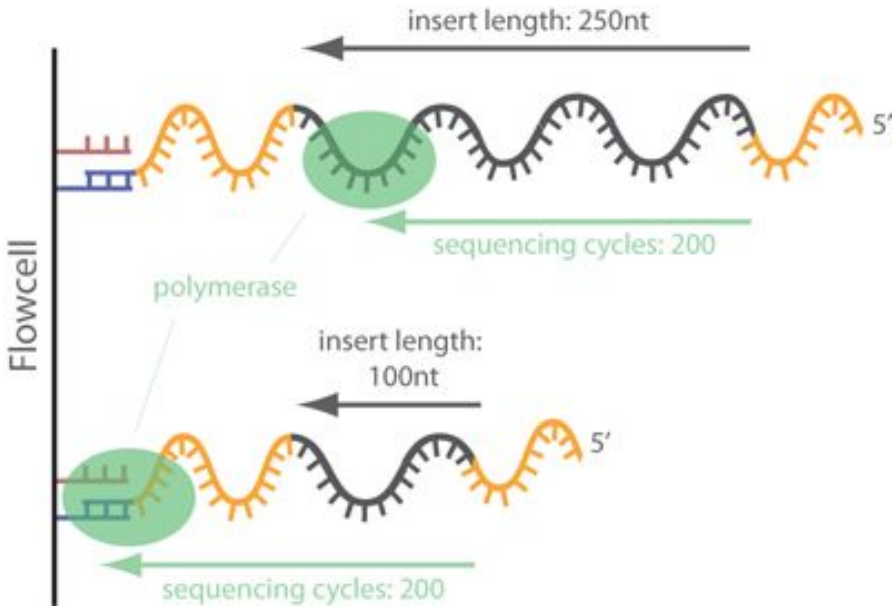
- Reads duplicates
- Assembly → *Hélène Touzet*

Reads cleaning

- Cut adaptators at read ends



https://www.ecseq.com/support/ngs/trimming-adapter-sequences-is-it-necessary

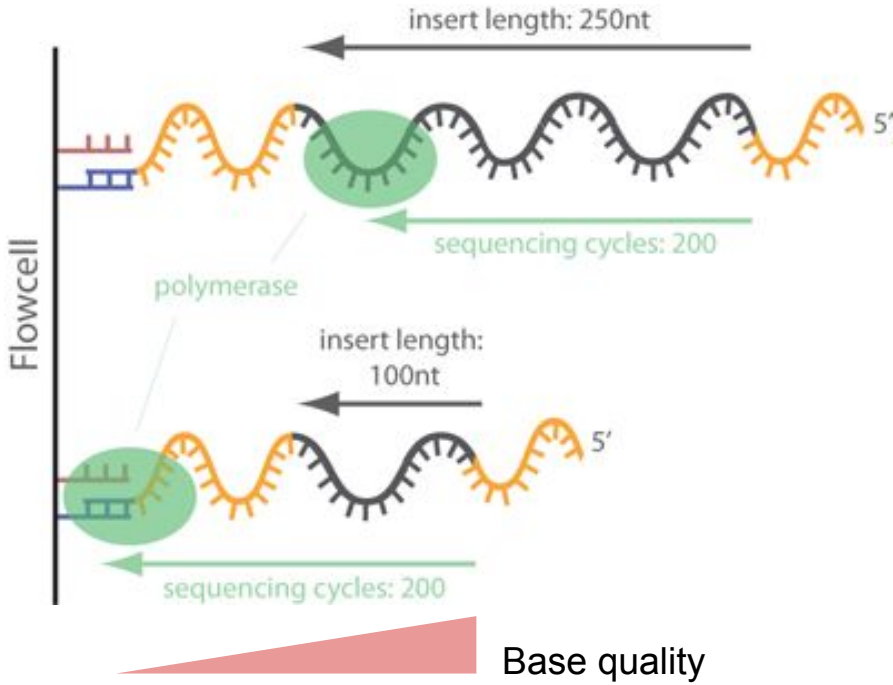
Reads cleaning

- Cut adaptators at read ends

- Trimming : cut read ends (5' ou 3')
  - Fixed number of bases
  - Individual base quality
  - Mean quality of bases in a sliding window

- Filtering : remove read
  - Size criteria (example < 60bp)
  - Mean base quality for all bases criteria (example < 25)

Reads cleaning



- Cut adaptators at read ends

- Trimming : cut read ends (5' ou 3')
  - Fixed number of bases
  - Individual base quality
  - Mean quality of bases in a sliding window

- Filtering : remove read
  - Size criteria (example < 60bp)
  - Mean base quality for all bases criteria (example < 25)

Reads cleaning example

Tool: Trimmomatic

01
- Clean adaptators

02
- Trimming 5' et 3' on base quality (> 3)

03
- Trimming using sliding window (4 bases, Q < 20)

04
- Filtering on mean read quality (Q < 25)

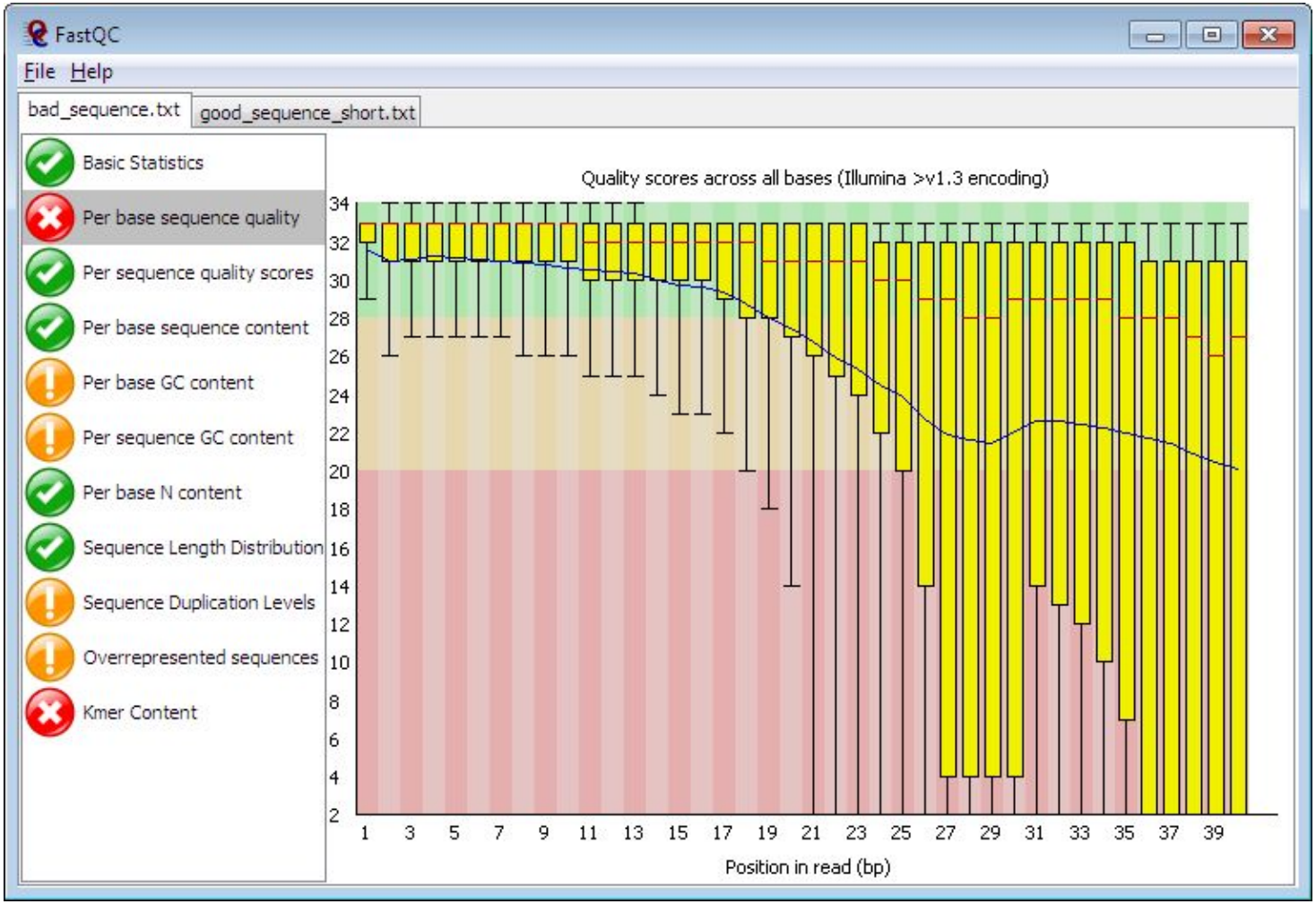
05
- Filtering on read size (taille < 20)

Source : Erwan Core, 5ème Ecole de bioinformatique AVIESAN-IFB 2016 https://www.france-bioinformatique.fr/fr/evenements/EBA2016

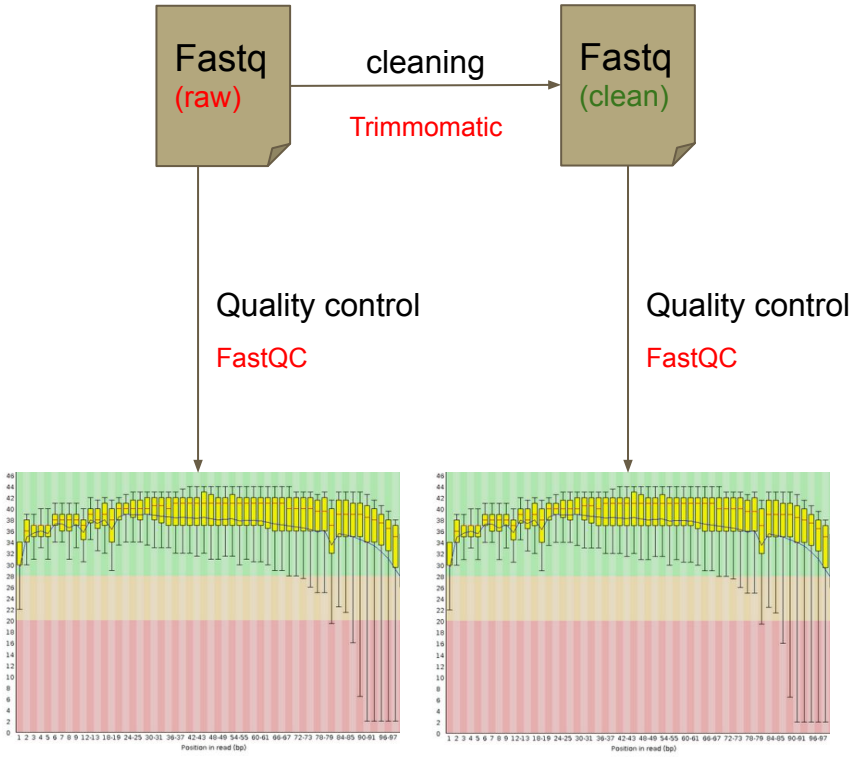
Bolger, A. M. and Lohse, M. and Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina sequence data. In Bioinformatics, 30 (15), pp. 2114–2120

Reads quality control (FastQC)

FastQC aims to provide a simple way to do some quality control checks on raw sequence data coming from high throughput sequencing



Workflow


Fastq
(raw)
cleaning
Trimmomatic
Fastq
(clean)
Quality control
FastQC
Quality control
FastQC