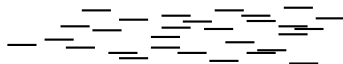# To map or not to map?

Formation RNA-Seq – Bilille

Mikaël Salson
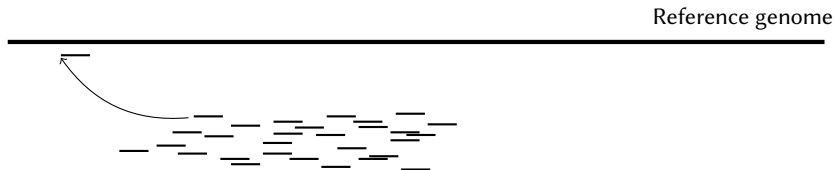
mikael.salson@univ-lille.fr

# RNA-Seq read mapping
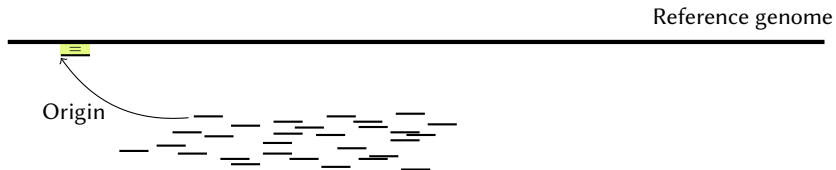
Reference genome

# RNA-Seq read mapping

Reference genome

# RNA-Seq read mapping

Reference genome

Origin

# RNA-Seq read mapping



Reference genome

Origin

# RNA-Seq read mapping

# RNA-Seq read mapping

# RNA-Seq read mapping

# RNA-Seq read mapping

# RNA-Seq read mapping

# RNA-Seq read mapping
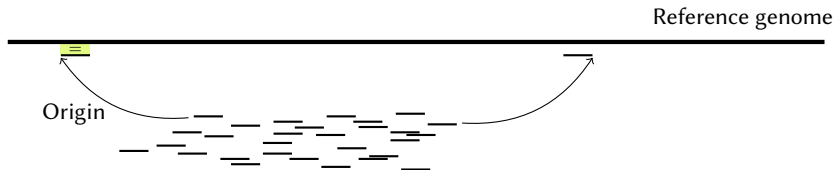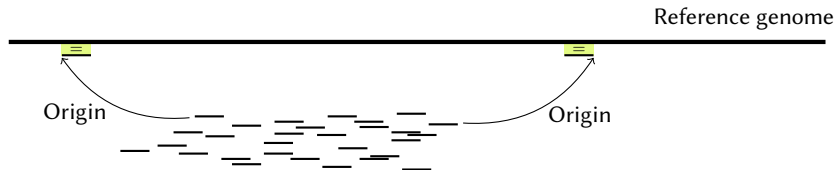


intron

Reference genome

Origin

Origin

# RNA-Seq read mapping
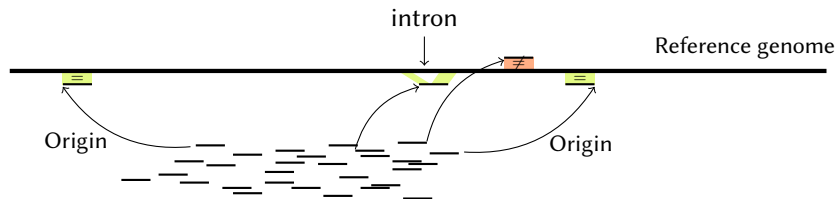
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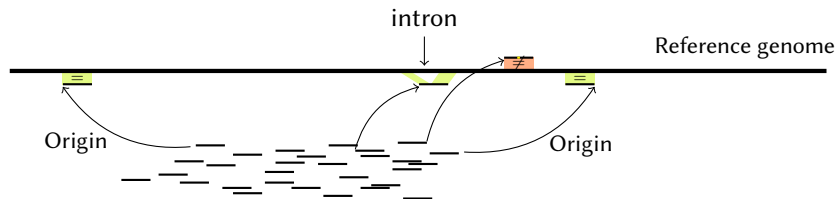
# RNA-Seq read mapping
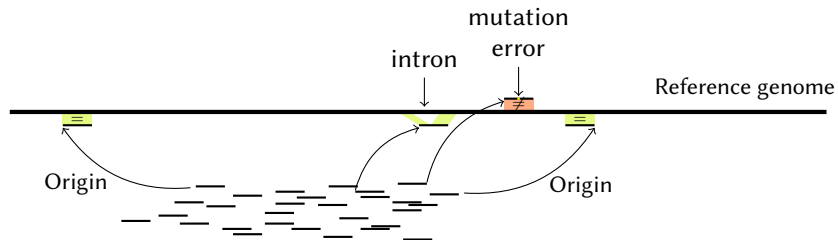
# RNA-Seq read mapping
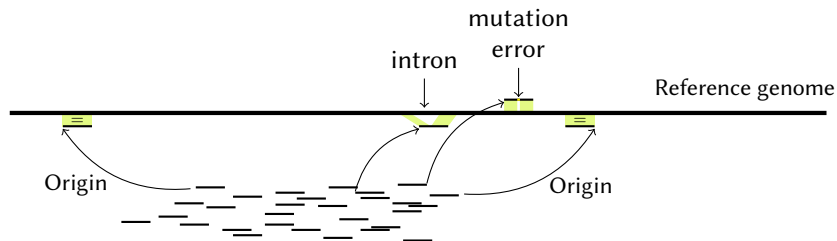
# RNA-Seq read mapping
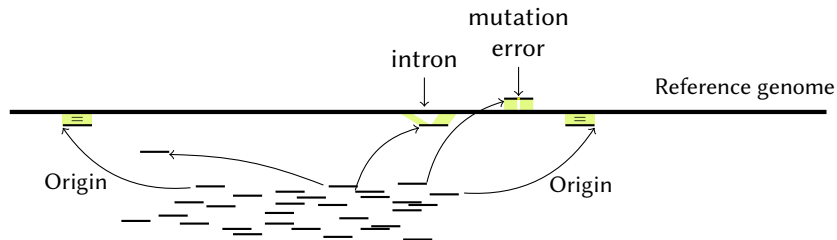
# RNA-Seq read mapping
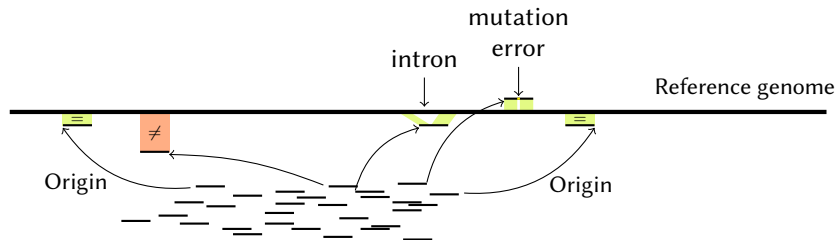
# RNA-Seq read mapping

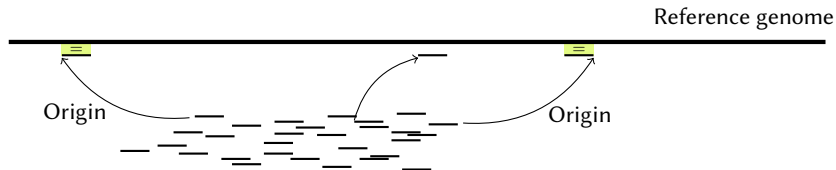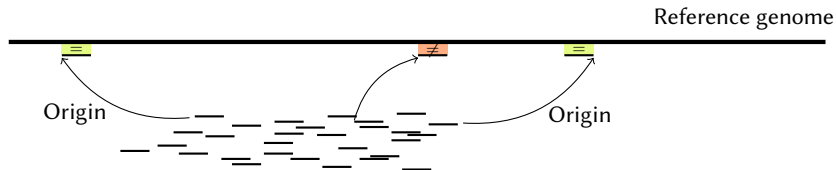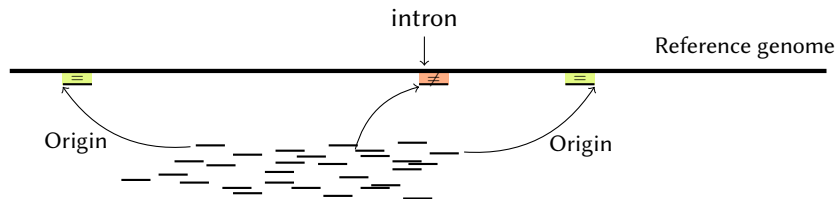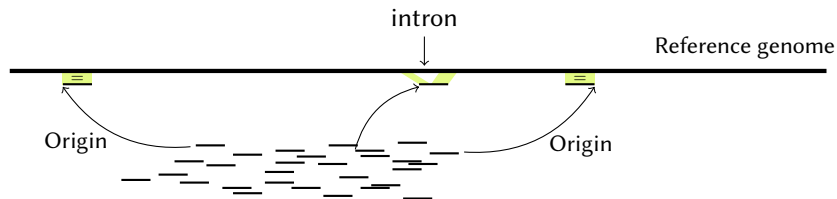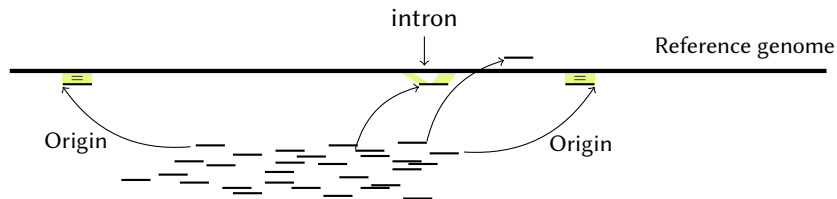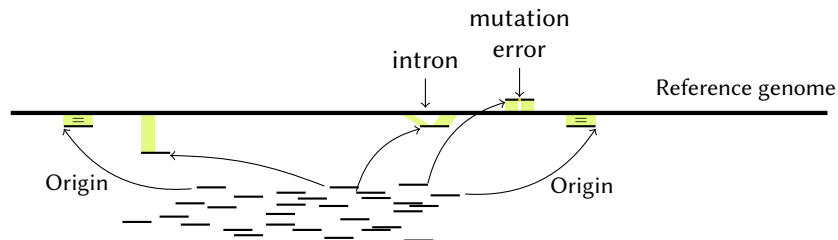# RNA-Seq read mapping
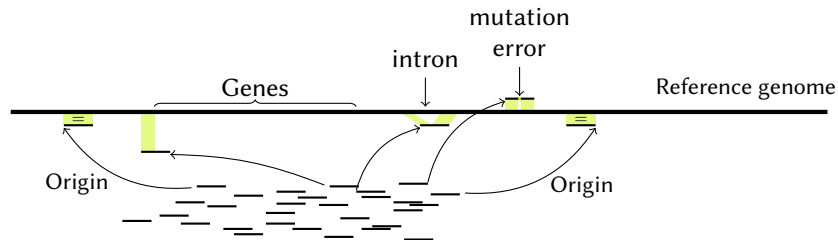
# RNA-Seq read mapping
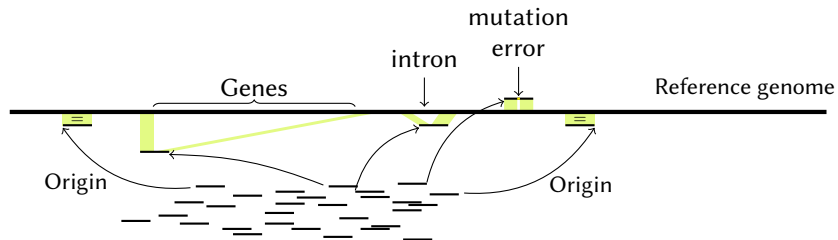
# RNA-Seq read mapping
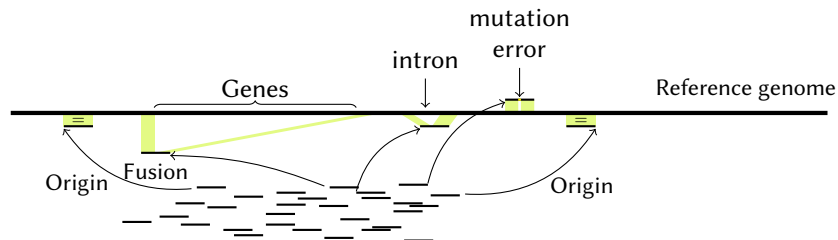
# Split reads don't align contiguously to the genome

Reads

# Split reads don't align contiguously to the genome
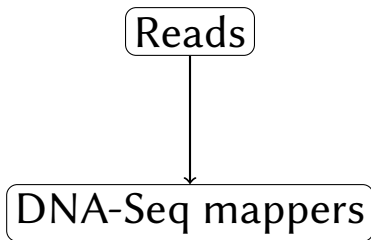
# Split reads don't align contiguously to the genome
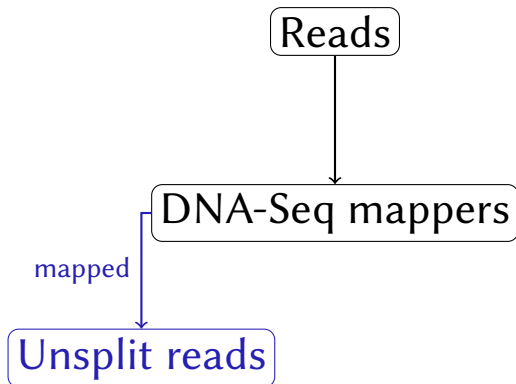
# Split reads don't align contiguously to the genome
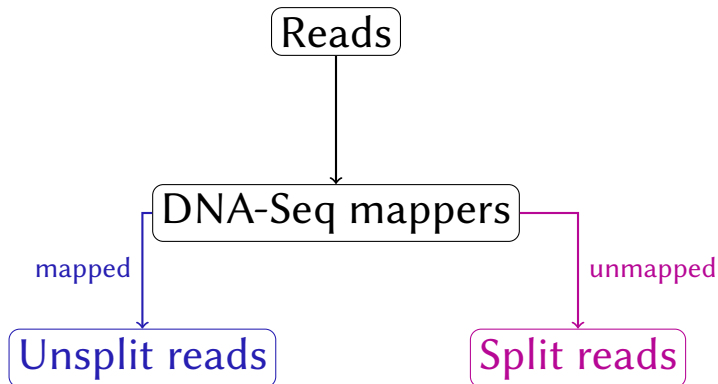
# Split reads don't align contiguously to the genome
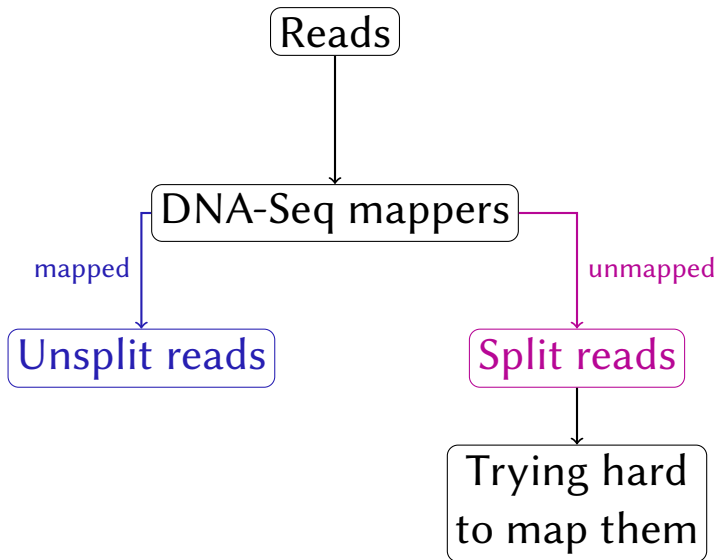
# Mapping split reads by… splitting them – TopHat2



© Tophat2: Kim et al, 2013

# Mapping all reads by splitting them – HISAT2, STAR



| e1 | e2 |

mismatch — x
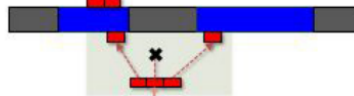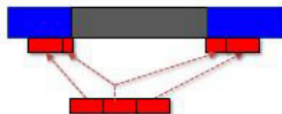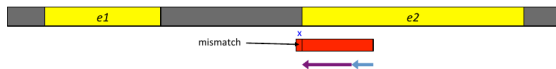
Local FM index for chr22 from 24,417,280 to 24,482,559
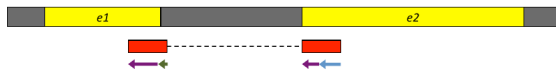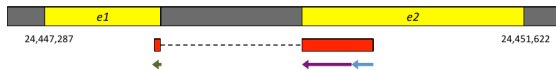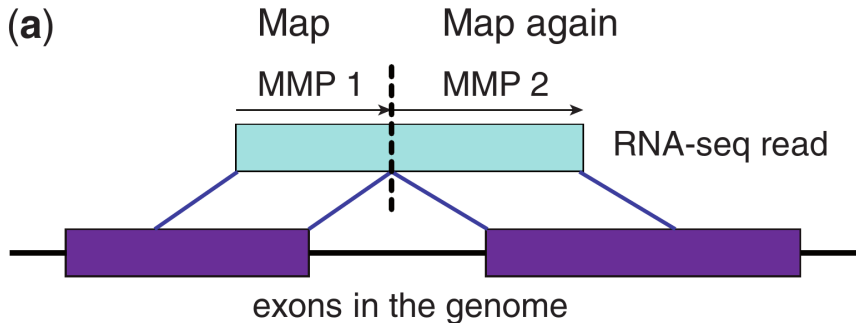
| e1 | e2 |

24,447,287    24,451,622

| e1 | e2 |

© HISAT: Kim, Langmead, Salzberg, Nat. Methods, 2015

# Mapping all reads by splitting them – HISAT2, STAR



(**a**)

Map — MMP 1 | Map again — MMP 2

RNA-seq read

exons in the genome

© STAR: Dobin et al, Bioinformatics, 2013

# Mapping methods

| | |
|---:|:---|
| **TopHat2** | Exact contiguous fixed-lengh |
| **HISAT** | Maximal mappable suffix |
| **STAR** | Maximal mappable prefix |

# Mapping methods

| | |
|---|---|
| **TopHat2** | Exact contiguous fixed-lengh |
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| **STAR** | Maximal mappable prefix |

# Indexing methods

| | |
|---|---|
| **TopHat2** | FM-index |
| **HISAT** | Multiple FM-indices |
| **STAR** | Suffix Array |

# Indexing methods

$$T = \overset{0}{C}\,\overset{1}{T}\,\overset{2}{A}\,\overset{3}{G}\,\overset{4}{T}\,\overset{5}{T}\,\overset{6}{A}\,\overset{7}{G}\,\overset{8}{\$}$$

# Indexing methods

$$T = \underset{\substack{0\ 1\ 2\ 3\ 4\ 5\ 6\ 7\ 8}}{\text{C T A G T T A G \$}}$$

| TS | 8 | 6 | 2 | 0 | 7 | 3 | 5 | 1 | 4 |
|----|---|---|---|---|---|---|---|---|---|
|    | \$ | A | A | C | G | G | T | T | T |
|    |    | G | G | T | \$ | T | A | A | T |
|    |    | \$ | T | A |   | T | G | G | A |
|    |    |   | T | G |   | A | \$ | T | G |
|    |    |   | A | T |   | G |   | T | \$ |
|    |    |   | G | T |   | \$ |   | A |   |
|    |    |   | \$ | A |   |   |   | G |   |
|    |    |   |   | G |   |   |   | \$ |   |

# Indexing methods

$$T = \overset{0\ \ 1\ \ 2\ \ 3\ \ 4\ \ 5\ \ 6\ \ 7\ \ 8}{\text{C T A G T T A G \$}}$$

| TS | 8 | 6 | 2 | 0 | 7 | 3 | 5 | 1 | 4 |
|----|---|---|---|---|---|---|---|---|---|
|    | \$ | A | A | C | G | G | T | T | T |
|    | C | G | G | T | \$ | T | A | A | T |
|    | T | \$ | T | A | C | T | G | G | A |
|    | A | C | T | G | T | A | \$ | T | G |
|    | G | T | A | T | A | G | C | T | \$ |
|    | T | A | G | T | G | \$ | T | A | C |
|    | T | G | \$ | A | T | C | A | G | T |
|    | A | T | C | G | T | T | G | \$ | A |
|    | G | T | T | \$ | A | A | T | C | G |

# Indexing methods

$$T = \overset{0}{C} \overset{1}{T} \overset{2}{A} \overset{3}{G} \overset{4}{T} \overset{5}{T} \overset{6}{A} \overset{7}{G} \overset{8}{\$}$$

| TS | 8 | 6 | 2 | 0 | 7 | 3 | 5 | 1 | 4 |
|----|---|---|---|---|---|---|---|---|---|
| | $ | A | A | C | G | G | T | T | T |
| | C | G | G | T | $ | T | A | A | T |
| | T | $ | T | A | C | T | G | G | A |
| | A | C | T | G | T | A | $ | T | G |
| | G | T | A | T | A | G | C | T | $ |
| | T | A | G | T | G | $ | T | A | C |
| | T | G | $ | A | T | C | A | G | T |
| | A | T | C | G | T | T | G | $ | A |
| | **G** | **T** | **T** | **$** | **A** | **A** | **T** | **C** | **G** |

Burrows-Wheeler Transform

# What approach is the best? (slide courtesy of J. Audoux)



NATURE METHODS | ANALYSIS

## Simulation-based comprehensive benchmarking of RNA-seq aligners

Giacomo Baruzzo, Katharina E Hayer, Eun Ji Kim, Barbara Di Camillo, Garret A FitzGerald & Gregory R Grant

METHOD    OPEN ACCESS

## A benchmark for RNA-seq quantification pipelines

Mingxiang Teng, Michael I. Love, Carrie A. Davis, Sarah Djebali, Alexander Dobin, Brenton R. Graveley, Sheng Li, Christopher E. Mason, Sara Olson, Dmitri Pervouchine, Cricket A. Sloan, Xintao Wei, Lijun Zhan and Rafael A. Irizarry

NATURE METHODS | ANALYSIS   OPEN

## Systematic evaluation of spliced alignment programs for RNA-seq data

Pär G Engström, Tamara Steijger, Botond Sipos, Gregory R Grant, RGASP Consortium, Gunnar Rätsch, Nick Goldman, Tim J Hubb, Roderic Guigó & Paul Bertone

Article | OPEN

## Comparative assessment of methods for the fusion transcripts detection from RNA-Seq data

Am J Hum Genet. 2013 Oct 3; 93(4): 641–651.   PMCID: PMC3
doi: 10.1016/j.ajhg.2013.08.008
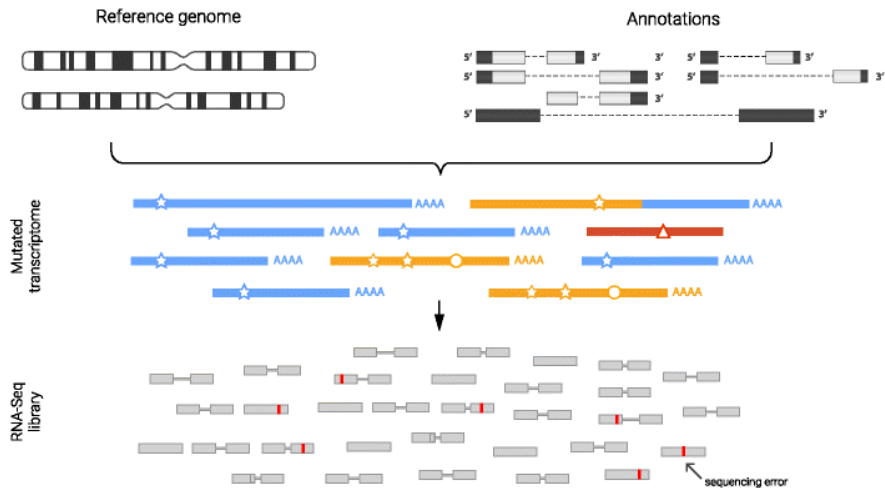
## Reliable Identification of Genomic Variants from RNA-Seq Data

Robert Piskol,[1] Gokul Ramaswami,[1] and Jin Billy Li[1,*]

Author information ► Article notes ► Copyright and License information ►

© Jérôme Audoux

# Benchmarking RNA-Seq aligners

Audoux *et al*, BMC Bioinformatics, 2017
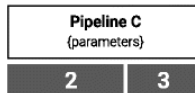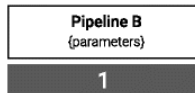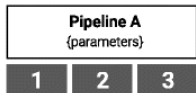


By Jérôme Audoux

# Benchmarking RNA-Seq aligners
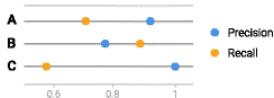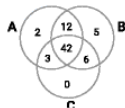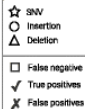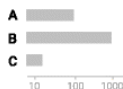
*By Jérôme Audoux*

# Sensitivity/accuracy of read mappers



### 160M 150bp reads from GRCh38
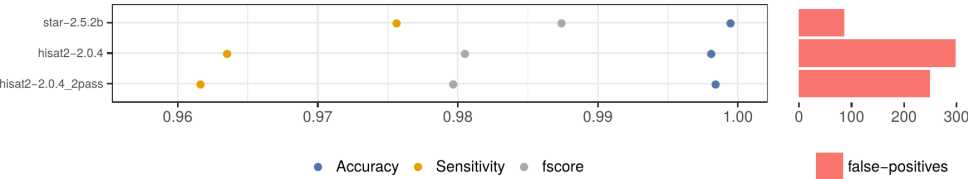
# STAR offers the best trade-off for splice detection



Splicing

160M 150bp reads from GRCh38
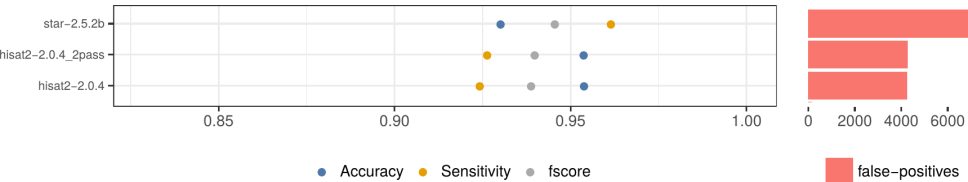
By Jérôme Audoux

# Space/time for read mappers
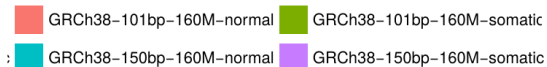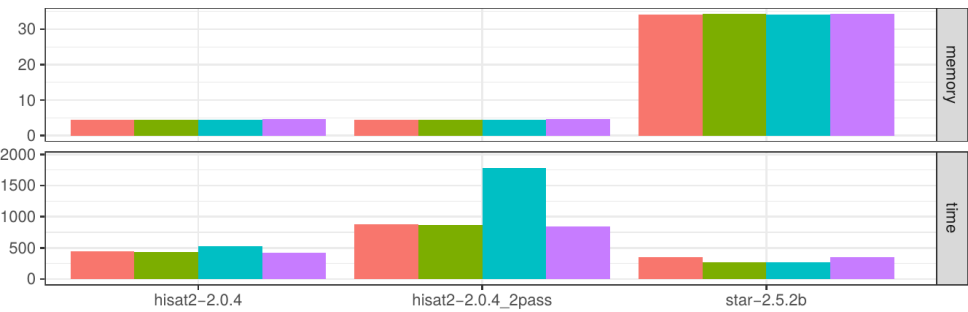


By Jérôme Audoux

# Many people uses TopHat2

# Many people uses TopHat2

## but don't

# but don't

## On TopHat2 website (since Feb 2016) ⧉

TopHat2 « *is now largely superseded by HISAT2 which provides the same core functionality (i.e. spliced alignment of RNA-Seq reads), in a more accurate and **much more efficient** way* » .

# Do you really need to map reads?

Does it matter to have a base pair precision
location for hundreds of millions of reads?

## Quantifying transcripts may not require alignment

**Kallisto**

Bray et al, Nat. Biotechnology, 2016

**Salmon**

Patro et al, Nat. Methods, 2017

# Quantifying transcripts may not require alignment

### Kallisto
Bray et al, Nat. Biotechnology, 2016 ⌕

### Salmon
Patro et al, Nat. Methods, 2017 ⌕

Two orders of magnitude faster than TopHat+Cufflinks

# How to quantify without aligning?



Transcripts

Read

Rest of the orange exon is
*uninformative* — this junction
is the *next informative
position*.

© *Rob Patro (Salmon)*

# How to quantify without aligning?



© Rob Patro (Salmon)
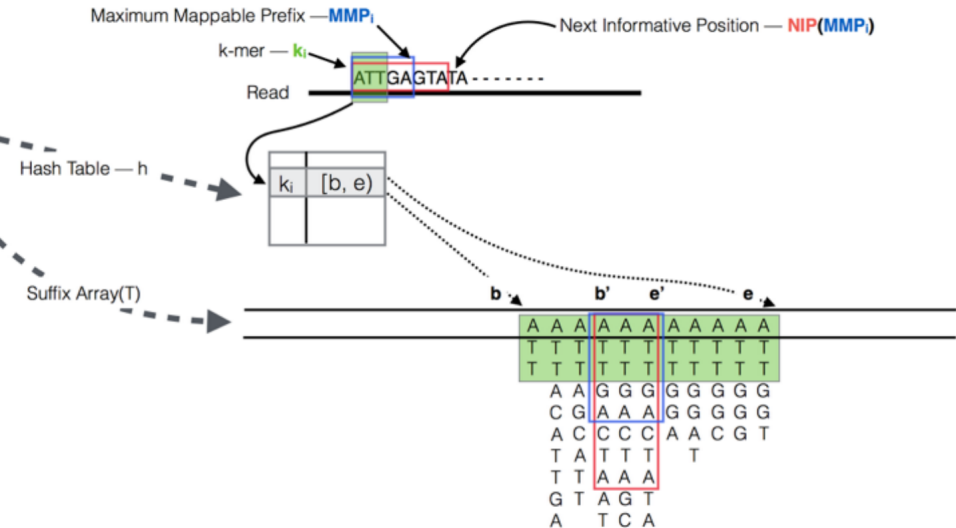
Ultra fast methods with good results...

# Ultra fast methods with good results…

« *With the exception of the underperforming Flux Capacitor and eXpress, we found that the other algorithms performed similarly.* »

Teng et al, Genome Biology, 2016

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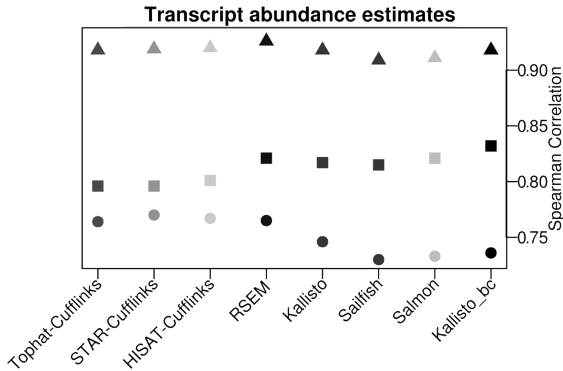


Germain et al, Nucleic Acid Research, 2016

# Ultra fast methods with good results…

« *With the exception of the underperforming Flux Capacitor and eXpress, we found that the other algorithms performed similarly.* »

Teng et al, Genome Biology, 2016 ↗

« *It is particularly noteworthy that Salmon, which (like Sailfish and Kallisto) bypasses traditional alignment and thereby quantifies a single sample in a matter of minutes, had a comparable performance to Cufflinks and RSEM. Importantly, we confirmed these results using a variety of assays on both empirical and simulated data.* »

Germain et al, Nucleic Acid Research, 2016 ↗

# Good performances may not hold true for all the data
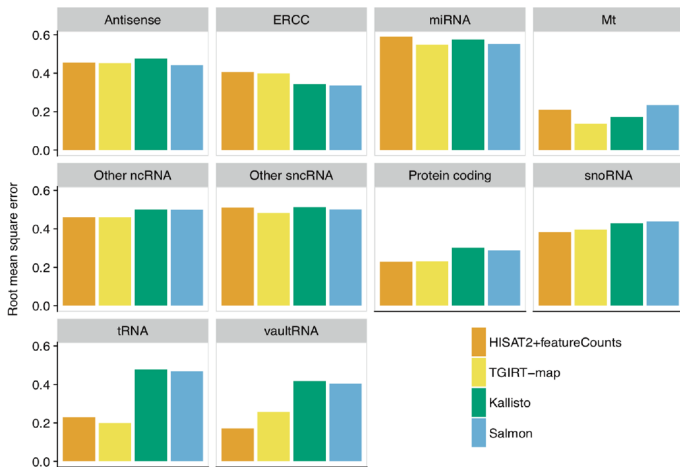
# Good performances may not hold true for all the data
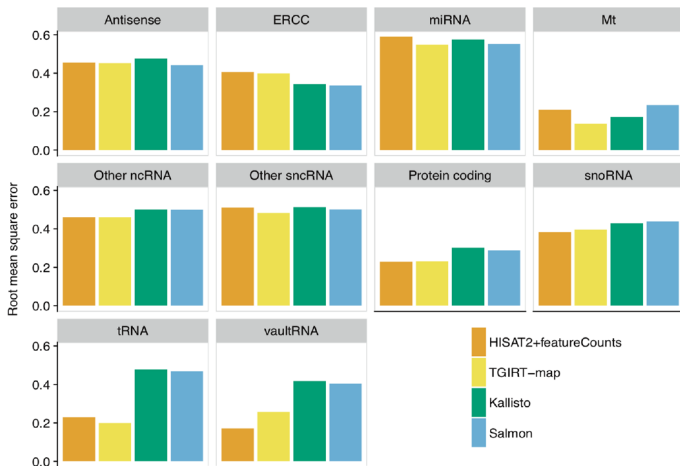
« *We have found that alignment-based tools were more accurate in quantifying lowly-expressed or small genes.* »

Wu *et al*, BMC Genomics, 2018

# Where the differences come from?

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1. Alignement vs pseudo-alignment

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2. Genome reference vs transcriptome reference
   see Srivastava *et al*, 2020 ⬀

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1. Alignement vs pseudo-alignment

2. Genome reference vs transcriptome reference
   see Srivastava *et al*, 2020 ⬈

3. Quantification method

# When a read maps at multiple loci,
# what transcript/gene should be counted?

See Deschamps-Francoeur *et al*, 2020 (thanks Pierre!)

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See Deschamps-Francoeur *et al*, 2020 ⌖ (thanks Pierre!)

► None
  (*eg.* HTSeq-count, STAR genecount, featureCounts)

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▶ Split counts evenly
  (*eg.* Cufflinks, featureCounts (with an option))

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▶ Rescue based on single mapping reads
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▶ Split counts evenly
  (*eg*. Cufflinks, featureCounts (with an option))

▶ Rescue based on single mapping reads
  (*eg*. Cufflinks (with an option))

▶ Expectation maximization
  (*eg*. RSEM, Salmon, Kallisto)

**High number of citations $\neq$ Best software**

**High number of citations $\neq$ Best software**

**Alignment isn't an end in itself**

**High number of citations $\neq$ Best software**

**Alignment isn't an end in itself**

**Alignment-free methods may be suitable for you**